

Rett syndrome: insights into genetic, molecular and circuit mechanisms

Jacque P. K. Ip¹, Nikolaos Mellios² and Mriganka Sur^{1*}

Abstract | Rett syndrome (RTT) is a severe neurological disorder caused by mutations in the gene encoding methyl-CpG-binding protein 2 (MeCP2). Almost two decades of research into RTT have greatly advanced our understanding of the function and regulation of the multifunctional protein MeCP2. Here, we review recent advances in understanding how loss of MeCP2 impacts different stages of brain development, discuss recent findings demonstrating the molecular role of MeCP2 as a transcriptional repressor, assess primary and secondary effects of MeCP2 loss and examine how loss of MeCP2 can result in an imbalance of neuronal excitation and inhibition at the circuit level along with dysregulation of activity-dependent mechanisms. These factors present challenges to the search for mechanism-based therapeutics for RTT and suggest specific approaches that may be more effective than others.

Classic RTT

The most common form of RTT. Several atypical forms of RTT have been described. The classic and atypical forms may differ by their symptoms or by the specific gene mutation.

Random X chromosome inactivation

Random inactivation of one X chromosome in early embryonic cells in females.

Somatic mosaics

The presence of two populations of cells with different genotypes (mutation-positive and mutation-negative) in one individual.

Rett syndrome (RTT; OMIM identifier #312750), first reported by the Austrian physician Andreas Rett, is a devastating neurological disorder that affects brain development and function in females in approximately 1 in 10,000 live births^{1,2}. In 90–95% of patients diagnosed with classic RTT, the disease is caused by loss-of-function mutations in the X-linked *MECP2* gene, which encodes methyl-CpG binding protein 2 (MeCP2).

MECP2 gene mutations predominantly arise in germ cells from the paternal side³. Despite the hundreds of RTT-causing *MECP2* mutations that have been identified, eight ‘hotspot’ mutations (encoding the amino acid substitutions R106W, R133C, T158M, R168X, R255X, R270X, R294X and R306C) constitute more than 60% of documented cases⁴. RTT-causing mutations in *MECP2* in female patients lead to developmental regression, including a loss of speech and hand skills, after apparently normal development⁵. In females, random X chromosome inactivation results in somatic mosaics with normal and mutant *MECP2*. *MECP2* mutations in males usually lead to severe congenital encephalopathies and death within 2 years⁶.

MeCP2 was initially identified as a nuclear protein that binds methylated DNA⁷ and was therefore described as a transcriptional repressor. Later, it was described as a multifunctional modulator of gene expression and as a transcriptional regulator with activating or repressing actions depending on the molecular context⁸. Recent evidence points to a role of MeCP2 as a weak repressor but with prominent secondary effects, including post-transcriptional regulation of gene expression through microRNA (miRNA)-mediated mechanisms⁹.

A large body of research on RTT produced over two decades has greatly advanced our understanding of the functions and regulation of MeCP2 as well as the mechanisms underlying RTT pathophysiology. Furthermore, recent studies have offered new insights into the roles of MeCP2 at different developmental stages, MeCP2–DNA interactions, the importance of MeCP2 for excitation–inhibition (E/I) balance at the circuit level and the plasticity that underlies RTT. The purposes of this Review are to provide a synthesis of the field, to assess the primary and secondary molecular actions of MeCP2 and to discuss how loss of MeCP2 results in changes in circuit function that are associated with RTT.

Clinical features of RTT

RTT is characterized by apparently normal early development with subsequent regression. However, subtly visible symptoms, such as hypotonia, jerkiness in limb movements and limited social interaction (as reflected by lack of interest in others), are present during early infancy¹⁰. Stunted head growth between 2 and 4 months of age may be diagnosed as acquired microcephaly and is an early sign of neurologic involvement¹⁰. The main symptoms often begin at around 12–18 months and include arrested cognitive and motor development and loss of acquired verbal skills. In addition, normal hand function is often replaced with stereotyped repetitive hand movements^{10,11}. This developmental stagnation and regression of acquired skills is characteristic of RTT. Another clinical hallmark is ataxia, which often results in motor disability, rendering affected individuals wheelchair-bound by the teenage years. Autonomic dysfunction and respiratory problems, seizures, anxiety and orthopaedic

¹Department of Brain and Cognitive Sciences, Picower Institute for Learning and Memory, Massachusetts Institute of Technology, Cambridge, MA, USA.

²Department of Neurosciences, School of Medicine, The University of New Mexico, Albuquerque, NM, USA.

*e-mail: msur@mit.edu

<https://doi.org/10.1038/s41583-018-0006-3>

problems are also common in individuals with RTT^{10,11}. Impaired development of visual processing and reduced visually evoked cortical responses have been reported in some patients^{12,13}.

The disease eventually plateaus and stabilizes¹⁰. Individuals with RTT live into adulthood and often require total care. Individuals with RTT may transiently exhibit autistic-like behaviours, such as social withdrawal and avoidance of eye gaze, during the regression phase, but they often become more interactive and social with age¹⁴. Notably, RTT is no longer listed as an autism spectrum disorder (ASD) in the fifth edition of the *Diagnostic and Statistical Manual of Mental Disorders* (DSM-V)¹⁵. However, RTT patients who show autistic-like features may be described as having ASD associated with RTT.

Neural phenotypes of RTT

The gross observation of reduced brain volume in individuals with RTT is associated with abnormally small and densely packed neurons that are reduced in dendritic complexity and spine density¹⁶. The dendrites of pyramidal neurons in the motor and frontal cortices are considerably shorter than in non-RTT brains¹⁶. MRI data demonstrate selective reductions in the volume of the parietal and temporal lobes in female RTT patients, with preservation of the occipital cortex¹⁷.

Following the identification of *MECP2* as the principal genetic factor underlying RTT¹⁸, enormous effort has been focused on modelling the disease using mouse and other animal models (BOX 1) and human induced pluripotent stem cell (iPSC)-derived models (BOX 2).

MeCP2 loss affects different cell types in multiple brain regions. MeCP2 is ubiquitously expressed but is enriched in the brain. Brain-specific deletion of *Mecp2* (*Nestin-Mecp2*^{-/-}) in mice captures key phenotypes, including reduced movement, abnormal gait, hindlimb claspings, respiratory abnormalities and early lethality, that present in *Mecp2*-null mice^{19,20}. This finding highlights a crucial role for the specificity of MeCP2 in the brain.

Region-specific knockout in the brain further reveals diverse roles of MeCP2 across various brain regions. For example, deletion of *Mecp2* in the brainstem and spinal cord leads to abnormal heart rate and breathing patterns and early lethality^{21,22}; removal from dopaminergic and noradrenergic neurons induces motor incoordination²³; and removal from serotonergic neurons increases aggression²³. Evidently, the region-specific loss of *Mecp2* induces a subset of RTT phenotypes that correspond to the function of the targeted region. Furthermore, the loss of MeCP2 in all inhibitory neurons (in *Viaat-Mecp2*^{-/-} mice) strikingly reproduces severe forms of RTT phenotypes, highlighting the important contribution of inhibitory neurons and E/I balance to MeCP2-mediated functions in the brain²⁴. Deletion of *Mecp2* from astrocytes in mice has relatively minor phenotypic consequences, but restoration of MeCP2 in astrocytes in an otherwise MeCP2-deficient nervous system results in marked amelioration of phenotypes²⁵. Transplantation of wild-type microglia to a mouse model of RTT was reported to reverse symptoms²⁶; however, multiple attempts to reproduce this finding were not successful²⁷,

pointing to the complexity of non-neuronal, potentially secondary, effects of MeCP2. Microglia have been suggested to contribute to RTT pathogenesis by excessively eliminating presynaptic inputs in fully symptomatic *Mecp2*-null mice. However, microglial-specific deletion (in *Cx3cr1*^{CreER}-*Mecp2*^{-/-} mice) has minimal effects on microglial phagocytosis and synaptic loss, suggesting involvement of other cell types in this effect²⁸.

Loss of MeCP2 affects multiple developmental stages.

Consistent with studies of post-mortem RTT brains, several early studies of mouse models suggested RTT to be a disorder of delayed neuronal maturation and synapse formation. These mouse models revealed reductions in neuronal soma size, dendritic arborization, spine density and postsynaptic density protein 95 (PSD95) immunoreactivity as well as abnormal spine morphology^{29–31}. Such findings led to the belief that MeCP2 is predominantly involved in the maturation of neuronal function, through processes including synaptogenesis, synaptic plasticity and establishment of functional circuits. In line with the structural defects, alterations in synaptic transmission are observed at the circuit level in *Mecp2*-mutant mice. Loss of MeCP2 leads to changes in neuronal excitability and deficits in experience-dependent plasticity processes, such as homeostatic synaptic scaling, resulting in altered neuronal connectivity and shifts in E/I balance at the circuit level (see details below). These functional deficits result from a failure of activity-dependent feedback mechanisms that require MeCP2 to establish and maintain functional brain circuits during neuronal maturation and throughout life^{32,33}. However, increasing evidence also suggests that MeCP2 has diverse and distinct roles in all stages of brain development, including the earliest stages of neurogenesis and neuronal migration (FIG. 1).

Initial reports in mouse models reported low MeCP2 expression in neural progenitor cells (NPCs) — with little effect of MeCP2 deficiency in cultured neurospheres — and a progressive increase during embryonic and postnatal development^{34,35}. More recent evidence now indicates that loss of MeCP2 affects early developmental stages considerably. Clinical studies report subtle symptoms before 6 months of age^{10,36–38} and early developmental delays before the full manifestation of RTT^{36,39,40}. Male babies born with a mutant *MECP2* allele, although very rare owing to the predominantly paternal origin of *MECP2* mutations, display severe neurological deficits from birth, including disproportionately small frontal and temporal lobes characteristic of a prenatal pathogenesis⁶.

Diverse phenotypes of RTT are present during neurogenesis and neuronal migration, and during the later stages of synaptic and circuit development and plasticity^{41,42}. Immunohistochemistry studies comparing wild-type cells with MeCP2-deficient cells suggest that MeCP2 is expressed in the prenatal cortex in mouse⁴² and in human iPSC-derived NPCs⁴³. The neurogenic function of MeCP2 has been demonstrated in mouse NPC cultures: overexpression of MeCP2 in these cultures inhibited glial differentiation but promoted neuronal differentiation⁴⁴. Conversely, embryonic NPCs extracted at embryonic day 15.5 from *Mecp2*-null mice exhibit a more proliferative identity and morphological alterations

Hypotonia

A state of low muscle tone (the amount of tension or resistance to stretch in a muscle), often resulting in muscle weakness.

Box 1 | Translational insights from mice

Research into Rett syndrome (RTT) received a major impetus through the generation and wide availability of methyl-CpG-binding protein 2 (*Mecp2*)-mutant mice^{19,20}. *Mecp2*-null (*Mecp2*^{-/-}) male mice are used more often than female heterozygous (*Mecp2*^{+/-}) mice, as the former display early-onset, severe symptoms similar to those in patients with RTT; namely, breathing irregularities and motor abnormalities, along with early lethality^{19,20}. Female *Mecp2*^{+/-} mice, which provide a more accurate genetic representation of the disorder, should be used as well as *Mecp2*^{-/-} mice for preclinical studies for potential therapeutics¹¹², even though their neurological symptoms develop later and they show higher phenotypic variability (owing to X chromosome inactivation)¹⁵⁶.

Mouse models for common missense mutations (including those encoding T158A, T158M and R306C substitutions) recapitulate RTT-like phenotypes^{68,157,158}. T158A and T158M affect the methyl-CpG-binding domain (MBD), reduce MeCP2 protein stability and cause reductions in the amplitude of event response potentials in the auditory cortex at the circuit level^{157,158}. R306C is a mutation that affects the nuclear receptor co-repressor (NCoR)–silencing mediator for retinoic acid and thyroid hormone receptor (SMRT) interaction domain (NID) and abolishes the interaction of MeCP2 with the NCoR–SMRT complex^{68,69}. As common missense mutations are highly relevant to RTT pathology, these models are valuable tools for preclinical studies.

One limitation of transgenic mouse models is that the onset of neurological symptoms occurs much later in development than in humans. Moreover, RTT-related phenotypes may be less severe in rodents. Other factors, including genetic background and rearing conditions, should be considered when analysing expressed phenotypes. Furthermore, RTT models in other species, including rats and non-human primates, have been developed. These recapitulate key features of RTT and offer unique behavioural features, including regression of a learned psychomotor skill in rats¹⁵⁹ and unique eye-tracking measures in primates¹⁶⁰. Such models could enable cross-species validation in developing preclinical outcome measures in the future¹⁵⁹.

RTT animal models have helped to establish biomarkers to track disease progression and amelioration. Several behavioural assessments have been established to track the onset and severity of RTT in mouse models¹⁵⁶. Unbiased, quantitative biomarkers of brain function and disease progression are needed. A recent study used frontal alpha-band electroencephalogram asymmetry, an index of anxiety and depression, in humans to evaluate the effects of insulin-like growth factor 1 (IGF1) treatment¹⁶¹. Other measurable phenotypes have been suggested: *Mecp2*^{-/-} mice exhibit regression of visual acuity in adulthood (as opposed to progressive increase of acuity during normal development)¹⁰⁹, and *Mecp2*^{+/-} mice and individuals with RTT exhibit decreases in the amplitude of cortical visually evoked potentials (VEPs)¹³. Therefore, a standardized VEP analysis might be useful in assessing preclinical and clinical therapeutic outcomes. The ideal biomarker may combine behavioural and electrophysiological assessments.

Mouse models have provided the basis for preclinical studies and therapeutic strategies for RTT (BOX 3). For example, several signalling pathways, including the brain-derived neurotrophic factor (BDNF)¹⁶² and IGF1 (REF.²⁹) pathways, are altered in RTT mouse models and are currently being targeted for potential therapeutics¹⁶³. BDNF has important roles in synaptic maturation and plasticity. BDNF levels are reduced in *Mecp2*-mutant mice¹⁶⁴, and candidate agents that increase BDNF levels (namely, glatiramer acetate and fingolimod) are in clinical trials^{165,166} in RTT¹⁶³. IGF1 regulates the phosphoinositide 3-kinase (PI3K)–AKT–postsynaptic density protein 95 (PSD95) pathway, a major pathway of excitatory synaptic maturation that is deficient in RTT model mice^{29,84} and in individuals with RTT¹⁶¹. PSD95 is a highly abundant synaptic protein crucial for synaptic maturation and plasticity^{167,168} and is upregulated by BDNF¹⁶⁹ and IGF1 (REFS^{29,170}). PSD95 is deficient in *Mecp2*-null mice¹⁷¹ and is restored following systemic treatment with IGF1 (REFS^{29,134}). Indeed, IGF1 treatment has shown efficacy in a phase I/IIa trial¹⁶¹. NNZ-2566, a synthetic analogue of IGF1, has been claimed to improve breathing and motor behaviour in patients with RTT; NNZ-2566 will soon advance to phase III trials.

as early as 3 days in vitro⁴². In *Xenopus laevis*, MeCP2 regulates the transcriptional expression of the neuronal repressor gene *XHairy2a* (also known as *hes4*); depletion of MeCP2 protein and expression of a truncated form of MeCP2 (R168X) result in a decrease and an increase in the number of NPCs, respectively⁴⁵. In zebrafish, genetic deletion of *mecp2* inhibits neuronal differentiation, whereas *Mecp2* overexpression promotes differentiation⁴⁶. Similarly, measurement of the expression of NPC markers and early neuronal markers in the *Mecp2*-null mouse brain shows an increase in NPC markers and a decrease in neuronal markers, suggesting a possible imbalance in prenatal brain development⁴². Moreover, alterations in human neurogenesis and neuronal differentiation are observed in patient-derived iPSC cultures and cerebral organoids⁴¹. Even small deflections in the developmental programme may predispose the embryo to complex phenotypes later on. Such reports of an early developmental role for MeCP2 are particularly important when we consider treatment time points for RTT patients. Although postnatal re-expression of MeCP2 at later stages

in *Mecp2*-mutant mice partially rescues several RTT phenotypes, including lifespan^{47,48}, the above findings suggest that earlier treatment would increase its effectiveness.

Earlier and more recent evidence suggests that MeCP2 is required for the maintenance of mature neural networks⁴⁹. Removing MeCP2 during postnatal development or in adulthood leads to rapid progression of RTT-like symptoms, including lethality^{50,51}. Several synaptic proteins, including NMDA receptor (NMDAR) subunits and presynaptic vesicular glutamate transporter 1 (VGLUT1), are reduced upon postnatal removal of MeCP2 (REF.⁵⁰). Intriguingly, the levels of mRNA encoding these proteins remain unchanged, suggesting that MeCP2 probably regulates the levels of these proteins post-transcriptionally⁵⁰. Further confirming the requirement of MeCP2 in the adult brain, in vivo genome editing by injection of adeno-associated viral vector (AAV)–*Streptococcus pyogenes* Cas9 and guide RNA targeting *Mecp2* in adult mouse primary visual cortex (V1) depleted MeCP2 in 70% of the infected cells and led to altered visual responses, including reduced orientation selectivity and peak response⁵².

Guide RNA

A short synthetic RNA composed of a 20-nucleotide sequence, homologous to the gene of interest, and a Cas9 nuclease-recruiting sequence for CRISPR–Cas9 gene editing.

Box 2 | Modelling RTT in human cells

Intact human brains cannot be subjected to irreversible genetic manipulations or high-throughput drug screening, and human brain samples can be obtained only post-mortem and represent only the end point of the disorder. Hence, induced pluripotent stem cell (iPSC)-derived cell culture models of human brain development have been important for elucidating specific cellular aspects of disease pathology. Genome editing using transcription activator-like effector nuclease (TALEN) and CRISPR technologies, together with human neuronal cultures derived from people with Rett syndrome (RTT), has revealed structural, molecular and physiological RTT-related phenotypes during early developmental stages^{41,43,74,172–175}.

Isogenic controls are crucial for iPSC analyses. In RTT, neurons expressing only the non-mutated methyl-CpG-binding protein 2 (*MECP2*) allele can be generated from patients' cells using random X chromosome inactivation, such that controls share the same genetic background as the patients^{41,43}. Similar to RTT mouse models and post-mortem human patient samples, RTT iPSC-derived neurons demonstrate reductions in soma size, spine density, neuronal firing and activity-dependent signalling¹⁷³. In addition, these cells show various signs of impaired neuronal maturation, including reduced expression of the neuronal marker TUJ1 (neuron-specific class III β -tubulin) and excitatory and inhibitory neuronal markers, aberrant electrophysiological properties and deficits in activity-dependent transcription^{43,74,172,175}. Moreover, RTT iPSC-derived cell cultures show impaired neuronal differentiation and migration⁴¹, and differentiated astrocytes from RTT patients have adverse effects on the morphology and function of wild-type neurons¹⁷⁴.

Cerebral organoids are stem-cell-derived in vitro 3D structures, with a certain self-organizing capacity, that mimic, at least in part, in vivo brains. Although cerebral organoids currently can be used only to model early development of the human brain, several key features of in vivo brain organogenesis are recapitulated by in vitro organoids, which thus provide robust models for studying neurological disorders¹⁷⁶. RTT patient-derived cerebral organoids have recently been generated and exhibit abnormalities in neuronal proliferation and differentiation as well as reduced maturity — phenotypes also observed in human *MeCP2*-deficient monolayer cultures⁴¹. However, to avoid the variability driven by random X chromosome inactivation, cerebral organoids are generated primarily from cells that express only the mutant copy of *MECP2*; thus, the effect more closely mimics human male mutations, which are more severe. Therefore, it will be of great interest to generate cerebral organoids that more closely mimic female RTT brains, with heterozygous expression and random X inactivation.

One intriguing difference between mouse models and human RTT is their disease severity. The effects of *MeCP2* deficiency are milder in rodents than in humans: *MeCP2*^{-/-} mice survive for several weeks postnatally, and *MeCP2*^{+/-} mice show late symptom onset. This difference puts human iPSC-derived models in a unique position for studying RTT pathophysiology. With rapid advances in organoid-culturing strategies, cerebral organoids can develop over an extended period (more than 9 months), establishing late-developmental and even mature features, including dendritic spine formation and active neuronal networks¹⁷⁷. If or when cell-type-specific differentiation becomes a routine feature of these models, they and human-derived 2D cultures will be very useful for studying the interaction between excitatory and inhibitory neurons in the context of RTT¹⁷⁸. In the meantime, cerebral organoids are suitable models to investigate early-developmental events, including neurogenesis and migration. As a future direction for precision medicine, drugs could potentially be tested on neuronal cultures derived from an individual with RTT before deciding whether they can be administered to the patient¹⁷⁹.

Molecular effects

Since the discovery in 1999 that mutations in the *MECP2* gene were the underlying cause of RTT¹⁸, tremendous progress has been made in understanding the molecular functions of MeCP2. Interacting with DNA is an essential part of MeCP2 function, as MeCP2 is primarily a chromatin-bound nuclear protein that acts as an epigenetic reader. On the basis of functional studies, MeCP2 has been proposed to regulate gene transcription, chromatin compaction and miRNA processing⁵³. This section discusses the molecular effects of MeCP2 and how its loss contributes to neurological deficits in RTT.

Epigenetics of MeCP2–DNA interactions. MeCP2 was originally found to bind specifically to DNA that contains methylated cytosines in CpG dinucleotides (mCG), via its methyl-CpG-binding domain (MBD), thus helping to repress gene expression^{7,54,55}. However, it was recently reported that MeCP2 also interacts with methylated CA (mCA) sites⁵⁶, and another report further specified the binding of MeCP2 to methylated CAC (mCAC) trinucleotides³⁷. Intriguingly, mCA is more abundant in postmitotic neurons, accumulates throughout development and seems to promote transcriptional repression^{58,59}. The sites that harbour mCA modifications are

frequently found in long genes that have been shown to be preferentially expressed in RTT⁵⁹. mCA sites are concentrated within gene bodies, which further suggests that MeCP2 obstructs transcription within transcribed regions⁶⁰. A recent study suggested that mCA sites are methylated during early postnatal life by DNA (cytosine-5)-methyltransferase 3A (DNMT3A)⁶¹ and that activity-induced sites are less DNMT3A-bound and are thus less methylated. This mCA-mediated repressive effect of MeCP2 is believed to mediate cell-type-specific transcriptional fine-tuning⁶¹.

The ability of MeCP2 to interact with DNA bearing 5-hydroxymethylcytosine (5hmC) was previously controversial. Several independent studies now show that MeCP2 can bind to hydroxymethylated CA (hmCA) sites, but with a much lower affinity than to hydroxymethylated CG (hmCG) sites (see REF.⁵⁶ for a more detailed discussion). 5hmC is frequently associated with gene activation (possibly owing to its low binding affinity to MeCP2), whereas the binding of MeCP2 to mCG and mCA is associated with repression⁶²; specifically, 5hmC is enriched in genes that are actively expressed in neurons⁶². It is possible that the hmCG modification may result in 'functional demethylation' of mCG sites, as it may promote the detachment of MeCP2 and thus

Epigenetic reader

A protein that reads specific epigenetic marks on DNA.

Long genes

Genes with length greater than 100 kb.

Gene bodies

Regions of DNA from the transcription start site to the end of the transcript.

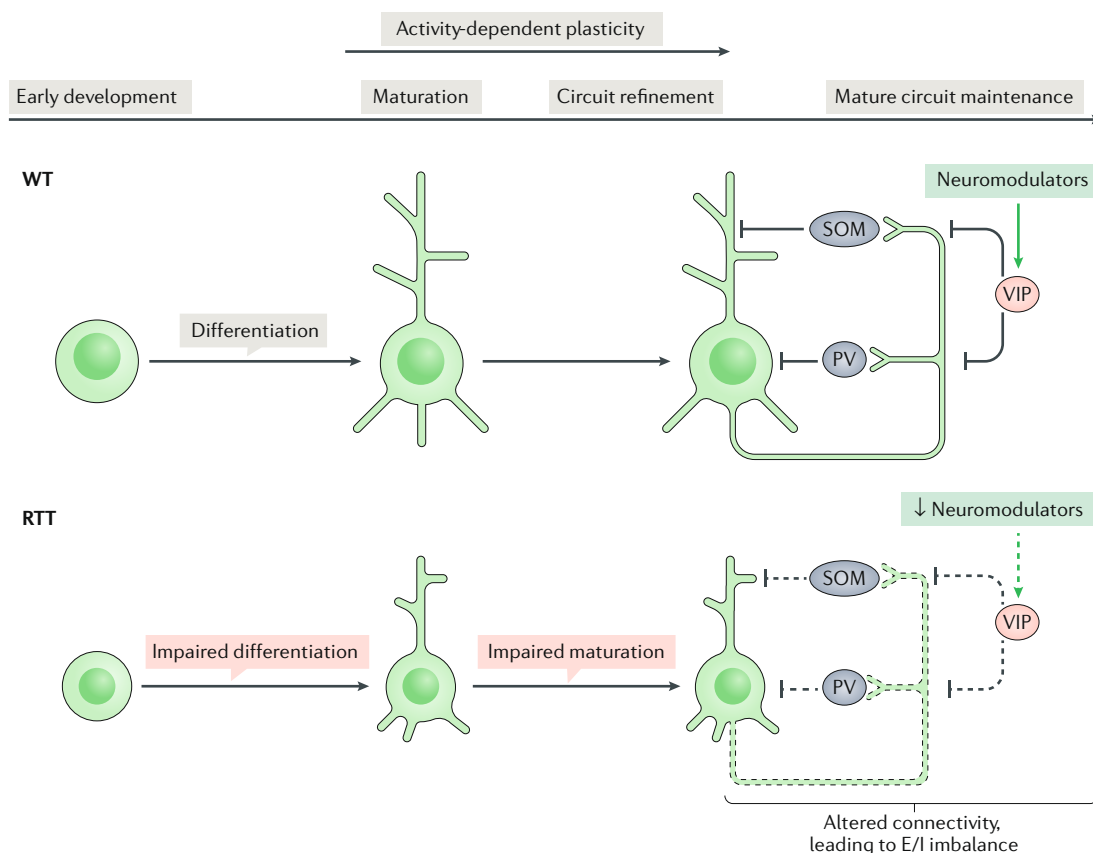


Fig. 1 | Effects of MeCP2 loss at different stages of brain development. Methyl-CpG-binding protein 2 (MeCP2) regulates multiple stages of brain development and function. Loss of MeCP2 results in impaired neurogenesis and differentiation in early development, abnormal neuronal maturation, reduced circuit connectivity and excitation–inhibition (E/I) imbalance — all of which potentially contribute to Rett syndrome (RTT) pathophysiology. Neural progenitor cells (left) differentiate into mature neurons through the development of dendrites and synapses and through experience-dependent and activity-dependent refinement. The proper development of microcircuits comprising pyramidal neurons and inhibitory neurons, including cells expressing somatostatin (SOM), parvalbumin (PV) or vasoactive intestinal peptide (VIP), is required for proper E/I balance. In RTT, the differentiation of neural progenitors is impaired at an early stage of development. At postnatal stages, RTT neurons show smaller soma sizes and underdeveloped dendritic arbors. Altered connectivity and reduced excitation and inhibition lead to E/I imbalance in RTT and RTT models; dashed lines represent altered connection strengths. WT, wild type.

achieve derepression of transcription⁶³. One model that synthesizes these recent findings on MeCP2–DNA interaction^{56,60,61,63} posits that activity and experience can modulate DNA methylation, through enzymatic actions, for the fine-tuning of MeCP2-regulated gene transcription (FIG. 2).

MeCP2 as a transcriptional regulator. A large body of data on the role of MeCP2 indicates that it is primarily a transcriptional repressor⁵³, although MeCP2 has also been suggested to act as a transcriptional activator⁸.

A role for MeCP2 in transcriptional repression was supported by the negative correlation between mCA levels and transcription of long genes⁵⁹. Moreover, two separate studies emphasized the proportional relationship between DNA methylation levels, MeCP2 binding on the genome and transcriptional repression^{57,60}. MeCP2 is highly abundant (there are an estimated 16 million molecules of MeCP2 per neuronal nucleus, compared with an estimated 32 million nucleosomes per nucleus), suggesting that it binds genome wide⁶⁴. Thus, converging lines of

evidence suggest that the repressive effects of MeCP2 are genome wide and positively correlated with the density of methylation of CG and CA sites in gene bodies⁶⁵.

Furthermore, microarray profiling and whole-cell RNA sequencing indicate that the repressive effect of MeCP2 is biased towards long genes larger than 100 kb, and expression of mutant MeCP2 leads to upregulation of the expression of long genes^{59,66}. More specifically, highly methylated long genes — and not lowly methylated long genes — are upregulated in *MECP2* mutants^{59,60}. A recent study analysing mice carrying specific RTT-associated *Mecp2* mutations (namely, those encoding the T158M and R106W substitutions) showed that gene expression changes are largely mutation-specific and cell-type-specific⁶⁷. Nuclear transcriptomes from forebrain neurons of these mutant mice reveal reductions in the nascent transcription of long genes and suggest a functional compensation for the reductions in long gene expression at the whole-cell level via post-transcriptional mechanisms⁶⁷. However, this study did not take into account the methylation status of the

Box 3 | Therapeutic strategies for RTT

There currently exist two major objectives in the development of therapeutic strategies: therapeutic targets downstream of methyl-CpG-binding protein 2 (MeCP2) and gene therapy. Postnatal restoration of MeCP2 in *Mecp2*-mutant mice rescues several Rett syndrome (RTT) phenotypes, emphasizing the reversibility of this devastating disorder^{47,48,180}. Several treatment strategies that are based on targeting signalling downstream of MeCP2, such as increasing brain-derived neurotrophic factor (BDNF) or insulin-like growth factor 1 (IGF1), have improved certain RTT features in preclinical tests and are in clinical trials^{161,163} (BOX 1). Given the diverse functional targets of MeCP2, it is likely that no single signalling pathway can explain disruptions resulting from MeCP2 dysfunction or loss. Rather, the development of novel treatment strategies must take into account not only the validity and specificity of the therapeutic target (that is, the mechanism of drug action and its relevance to MeCP2 mechanisms) but also the diversity of the effects of MeCP2 dysfunction or loss on different brain regions and cell types and during different developmental phases as well as on the time course of disease progression. Furthermore, therapeutic effectiveness is influenced by various factors, including the stratification and validity of patient populations and their match to preclinical drug validation models, drug dosage, trial onset and duration, effectiveness of biomarkers and choice of end points. Considering the temporal and regional features of RTT discussed in the main text, and the issues mentioned above, the most effective therapeutics will probably target mechanisms as directly, and as early, as possible. Current therapeutic strategies for RTT, including those in clinical trials, have been reviewed recently¹⁶³.

As inhibitory dysfunction is a major feature of RTT, its modulation is a feasible approach to a functional treatment for RTT. As excitation is also reduced in forebrain areas in *Mecp2*-mutant mice, enhancing excitation should also be considered for treatment strategies. Strikingly, deep brain stimulation (DBS) targeting the hippocampus rescues memory deficits in *Mecp2*-mutant mice^{181,182}, presumably by modulating the excitation and inhibition of local circuits¹⁸³. This finding opens a potential new avenue for RTT therapy. Mechanism-based approaches, such as IGF1 treatment (BOX 1), restore inhibition and excitation in *Mecp2*-mutant mice¹⁰¹. Ketamine, an NMDA receptor antagonist, improves specific functions, including forebrain activity (as indicated by FOS expression) and prepulse inhibition of the acoustic startle in *Mecp2*^{-f/y} and *Mecp2*^{-/-} mice, respectively¹⁰⁶. MeCP2-deficient mice and neurons derived from individuals with RTT show a lower K⁺/Cl⁻ exporter 2 (KCC2):Na⁺/K⁺/Cl⁻ symporter (NKCC1) ratio, leading to abnormal GABA-induced changes in membrane response and thus reduced inhibition. The US Food and Drug Administration (FDA)-approved NKCC1 antagonist bumetanide could thus be a potential candidate^{101,130}, although the development of molecules that enhance KCC2 function would be an important advance.

Owing to the complexity of MeCP2 dysfunction in RTT, gene therapy is possibly the most direct approach for treating the disorder. In principle, gene therapy would need to be attempted as early as possible, but the level of MeCP2 would have to be accurately titrated, as overdosage of MeCP2 or duplication of *MECP2* would lead to severe neurological consequences^{153–155}. Current evidence supports four potential gene therapy strategies^{11,163}. First, delivery of *Mecp2* gene using an adeno-associated virus serotype 9 (AAV9) vector through intravenous injection partially rescues RTT-like features in *Mecp2*-mutant mice¹⁸⁴. Second, molecules such as aminoglycoside antibiotics (for example, gentamycin) can enable ribosomes to read across MeCP2-encoding RNA despite nonsense mutations^{185,186} and could therefore potentially be used to target nonsense mutations that generate truncated MeCP2. Third, small molecules can activate normal *MECP2* alleles on the inactive X chromosome (Xi)¹⁸⁷. A recent study used a mixed-modality approach, combining brain-specific deletion of X-inactivation master regulator *Xist* and a small-molecule inhibitor of DNA methylation, to selectively reactivate Xi in mice in vivo¹⁸⁸. Fourth, gene editing using CRISPR–Cas9 techniques may be used to correct point mutations. As a complementary proof of principle, the CRISPR–Cas9 technique has been shown to be able to remove 70% of MeCP2 protein in the mouse hippocampus⁵². Site-directed RNA editing mediated by double-stranded RNA-specific editase 1 (ADAR2; also known as ADARB1) can correct disease-causing mutations affecting the mouse MeCP2 DNA-binding domain in vitro, repairing 72% of *Mecp2* mRNA¹⁸⁹. Although many technical barriers remain, the potential for gene therapy for RTT warrants further exploration.

long genes affected by MeCP2 loss, which is particularly important because other studies found that only highly methylated long genes are upregulated in *Mecp2*-knockout mice^{59,60}. Future studies will be required to further delineate the effects of various *MECP2* mutations on the transcriptional regulation of long genes in different neuronal cell types and to correlate these effects with the methylation status of long genes.

Considerable progress has recently been made in resolving the molecular mechanisms underlying the repressor role of MeCP2. MeCP2 has been reported to interact with the histone deacetylase (HDAC)–SIN3A co-repressor complex as well as with the HDAC3-containing nuclear receptor co-repressor (NCoR)–silencing mediator for retinoic acid and thyroid hormone receptor (SMRT) complex^{55,68}. The recruitment of the NCoR–SMRT complex has been particularly implicated in mediating the repressive effects of MeCP2 (REF. 68). The NCoR–SMRT

complex contains multiple subunits, including NCoR, SMRT, HDAC3, G protein pathway suppressor 2 (GPS2) and transducin β -like protein 1 (TBL1) or its paralogue TBL1-related protein 1 (TBLR1)⁶⁹. MeCP2 recruits the NCoR–SMRT complex by directly interacting with TBL1 and TBLR1 through its NCoR–SMRT interaction domain (NID) within its transcriptional repression domain (TRD)⁶⁹. In this model, the HDAC3-containing NCoR–SMRT complex is recruited by MeCP2 when MeCP2 is bound to DNA, and modulates chromatin structure by removing acetyl groups from histone lysine residues^{55,68} (FIG. 3). Notably, RTT-associated *Mecp2* mutations (such as R306C) that affect the NID of MeCP2 result in severe RTT phenotypes⁶⁸, and a recent study demonstrated that the MBD and NID of MeCP2 may be most phenotypically relevant for RTT⁷⁰. Thus, the primary function of MeCP2 is probably to connect methylated DNA with the NCoR–SMRT co-repressor complex. The binding of

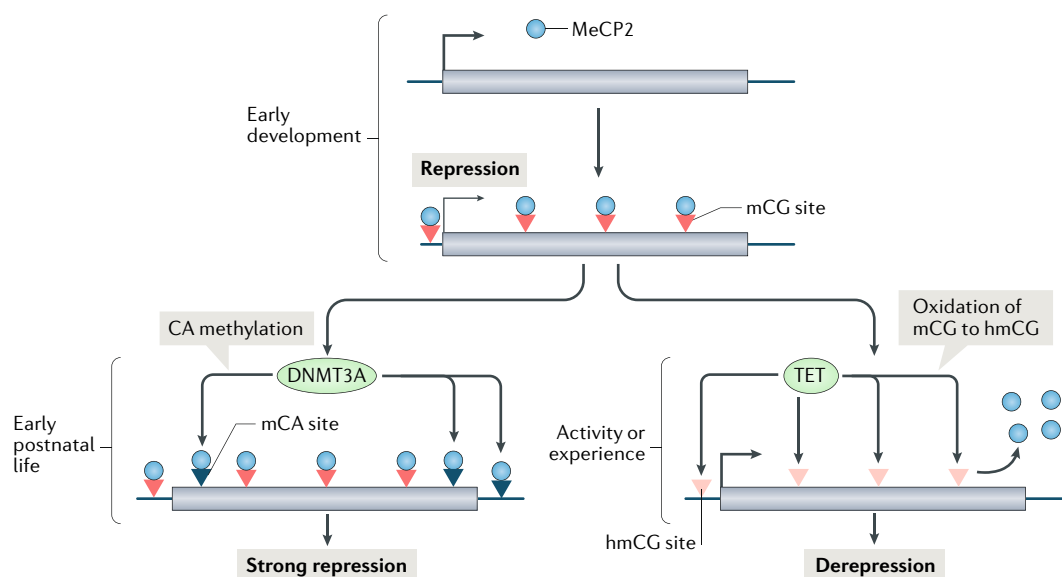


Fig. 2 | A model of MeCP2–DNA interactions. During early development, methyl-CpG-binding protein 2 (MeCP2) binds to genomic methylated CG (mCG) dinucleotide sequences to repress gene expression. During early postnatal life, DNA (cytosine-5)-methyltransferase 3A (DNMT3A) binds across transcribed regions of genes that show low expression and dictates DNA methylation at CA sequences (producing methylated CA (mCA) modifications)⁶¹. Once methylated, mCA is bound by MeCP2, leading to strong repression of gene transcription⁶¹. Passive ‘functional demethylation’ can occur through oxidation of mCG to hydroxymethylated CG (hmCG) by the ten-eleven translocation (TET) enzyme. As MeCP2 has a low affinity for hmCG, this modification results in the detachment of MeCP2, leading to derepression of transcription⁶³.

the NCoR–SMRT complex to MeCP2 is also regulated in an activity-dependent manner (see below) and is required for MeCP2-mediated gene repression^{68,71}. Although the relationship between methylation density and MeCP2 occupancy across the genome is well established, whether the MeCP2–NCoR–SMRT–HDAC3 complex is recruited in a density-dependent way, and is concentrated in the gene bodies of long genes, remains to be determined.

Notably, transcriptome analyses have revealed upregulation and downregulation of different genes in MeCP2-deficient mouse brains^{8,72,73}. As such, MeCP2 has been suggested to also act as a transcriptional activator and is thought to do so via its interaction with cAMP-responsive element-binding protein 1 (CREB1)⁸. However, no RTT mutation has yet been reported to interrupt MeCP2–CREB1 interaction. One possible explanation for the downregulation of a large numbers of genes in MeCP2-deficient neurons is that the transcriptional machinery does not function optimally, as reflected by the reduction of soma size and total RNA in RTT mouse models and in iPSC-derived neurons from individuals with RTT^{57,74}. This suggestion is consistent with the proposed function of MeCP2 as a multifunctional hub of cellular metabolism⁷⁴.

A recent study suggested that MeCP2 might act as a positive regulator for a subset of neuronal genes via its NID⁷⁵. At certain gene promoters, MeCP2 recruits HDAC3 to deacetylate the transcription factor forkhead box protein O3 (FOXO3) to positively regulate transcription (FIG. 3). In support of this mechanism, *Hdac3*-mutant mice (which show RTT-like phenotypes) and human iPSC-derived NPCs harbouring *MECP2*^{R306C} show mislocalization of FOXO3 (outside of gene promoters) and downregulation of target genes⁷⁵.

Nevertheless, this mechanism is surprising, as methylation density and MeCP2 occupancy are known to be low in transcription start sites, and MeCP2 generally binds within gene bodies^{57,76}. To reconcile these observations, one possibility is that the binding of MeCP2 to the gene body represses transcription, whereas its recruitment of HDAC3 to certain gene promoters positively regulates transcription through FOXO3 deacetylation⁷⁵.

An additional model proposes that MeCP2 directly interacts with DNA and thus modifies chromatin architecture globally^{64,77} (FIG. 3). *Mecp2*-null brains show global alterations in neuronal chromatin architecture, including increases in histone acetylation and elevated levels of linker histone H1 (REF.⁶⁴). As the NID contains a cluster of basic amino acids, it may act as a DNA-binding domain⁷⁸. In addition, MeCP2 has three AT hooks that could bend DNA and assist chromatin-structure remodelling⁷⁷.

Also relevant to a role for MeCP2 in maintaining chromatin structure is the interaction between MeCP2 and the transcription regulating helicase ATRX. MeCP2 and ATRX colocalize on pericentric heterochromatin, and ATRX is mislocalized in MeCP2-deficient neurons^{77,79}. The MeCP2–ATRX interaction may be important for the formation of higher-order chromatin loops⁷⁷. However, more direct evidence is required to demonstrate how MeCP2 may regulate chromatin structure, as well as the role of a loss of MeCP2–ATRX interactions in RTT pathology.

Interplay between MeCP2 and miRNAs. miRNAs are important for many developmental processes, including neurogenesis, brain maturation and synaptic plasticity^{80,81}. miRNAs have been reported to be dysregulated in the (total) brain, hippocampus and cortex of

Transcription start sites

The location where transcription starts at the 5′ end of a gene sequence. Promoters are located near the transcription start sites of genes.

AT hooks

DNA-binding motifs that bind specifically to AT-rich DNA.

Chromatin loops

When stretches of genomic sequence in the same chromosome are physically closer together than to the intervening sequences. These are involved in regulating chromatin organization and gene expression.

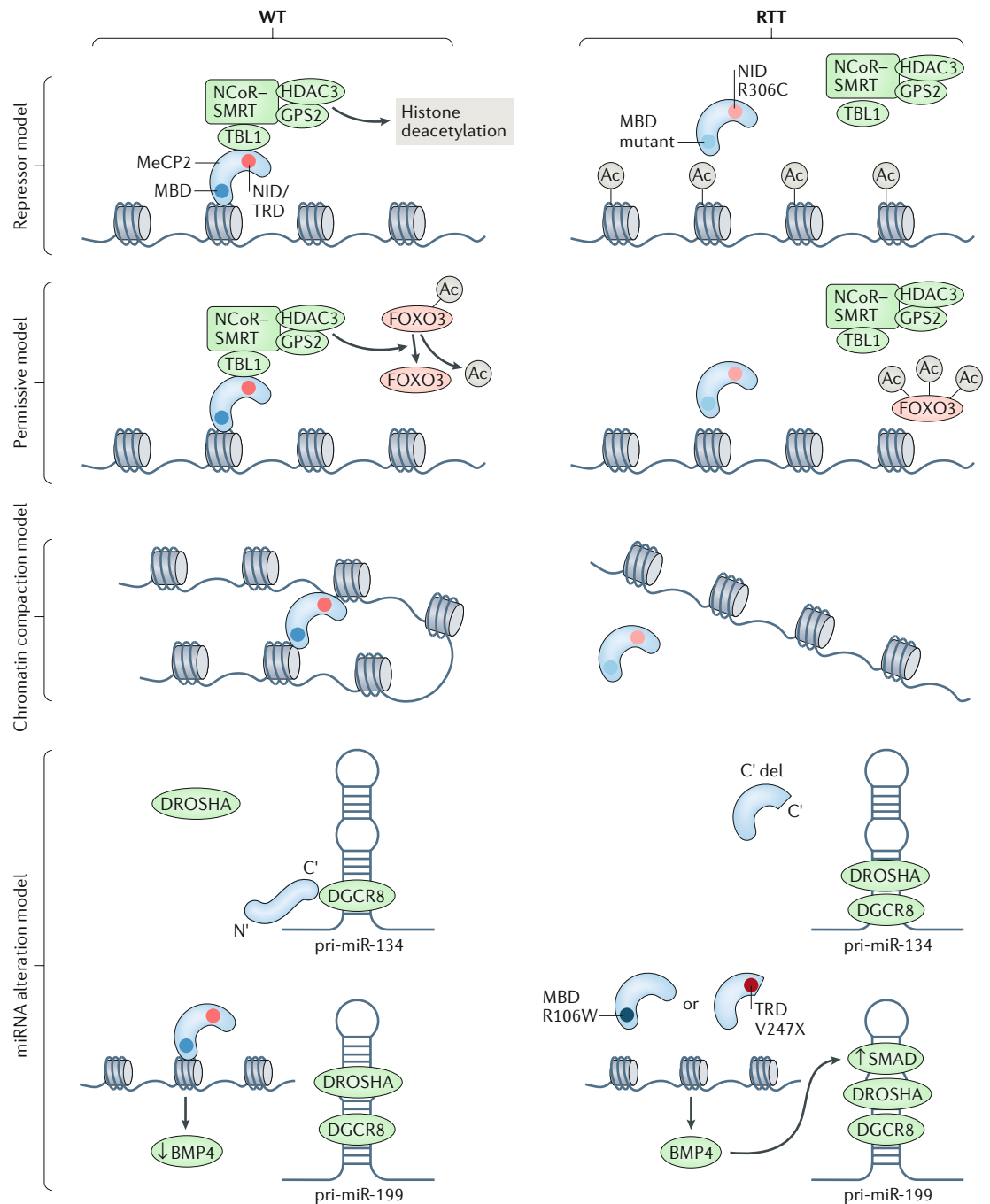


Fig. 3 | Proposed molecular effects of MeCP2 and their alterations in RTT. Methyl-CpG-binding protein 2 (MeCP2) has been proposed to recruit the nuclear receptor co-repressor (NCoR)–silencing mediator of retinoic acid (SMRT) complex to methylated DNA to repress transcription⁶⁸ or permit transcription⁷⁵ through the action of histone deacetylase 3 (HDAC3) depending on different genomic contexts. MeCP2 interacts directly with transducin β -like protein 1 (TBL1) in the NCoR–SMRT complex via its NCoR–SMRT interaction domain (NID)⁶⁹. This function is abolished in NID variants of MeCP2, such as R306C MeCP2. MeCP2 has also been suggested to compact chromatin structure through a basic amino acid cluster in its NID as well as via three AT hooks (not shown)⁷⁷. The carboxyl terminus of MeCP2 binds to the microprocessor complex protein DiGeorge syndrome critical region 8 (DGCR8) and inhibits ribonuclease 3 (DROSHA) from binding; thus, MeCP2 may inhibit the processing of pri-miR-134 (REF.⁹). In Rett syndrome (RTT) patients with *MECP2* mutations that change or truncate the C terminus (for example, the MeCP2 380 truncation), it is expected that MeCP2 binding to DGCR8 is abolished and DROSHA is free to bind and activate pri-miR-134 processing. Other methyl-CpG binding domain (MBD)-affecting or truncation-inducing mutations in *MECP2* also lead to upregulations of pri-miR-199 indirectly, via aberrant bone morphogenetic protein 4 (BMP4) and SMAD signalling⁴¹. Ac, acetyl group; C', carboxyl terminus; FOXO3, forkhead box protein O3; GPS2, G protein pathway suppressor 2; miRNA, microRNA; TRD, transcriptional repression domain; WT, wild type. Figure adapted from REF.⁵³, Macmillan Publishers Limited.

Mecp2-null mice^{9,82–85}; in mouse MeCP2-deficient neuronal cultures^{86–89}; and in iPSC-derived neuronal cultures and cerebral organoids generated from individuals with RTT⁴¹. The importance of miRNA–MeCP2 interactions in understanding RTT is being determined by studies of how MeCP2 loss alters miRNA expression^{9,41,86–89} and the different MeCP2 domains (discussed above) and RTT-associated *MECP2* mutations that could be implicated in miRNA alterations^{9,41}.

MeCP2 represses the transcription of primary miRNAs (pri-miRNAs), including miR-137, miR-15a, miR-184 and miR-7b, by binding to mCG sites in the gene promoter regions^{86–89}. MeCP2 deficiency results in increased pri-miRNA transcription, which leads to increased mature miRNA levels. Therefore, RTT-associated mutations in the genetic sequence encoding the MeCP2 MBD and TRD or NID are expected to result in increases in such MeCP2-regulated intergenic miRNAs. Furthermore, MeCP2 can directly inhibit miRNA expression by blocking pri-miRNA processing. MeCP2 may do this by binding to the microprocessor protein DiGeorge syndrome critical region 8 (DGCR8) via the carboxy-terminal domain⁹ and thus hindering the interaction of DGCR8 with the other main component of the microprocessor complex, ribonuclease 3 (DROSHA) (FIG. 3). This mode of action has been shown to lead to increased pri-miR-134 processing and mature miR-134 expression in the hippocampus of adult *Mecp2*-mutant mice⁹. In individuals with RTT who carry *MECP2* C-terminal mutations or truncations, the binding between MeCP2 and DGCR8 is expected to be abolished, leaving DROSHA free to bind and activate pri-miR-134 processing (FIG. 3). Intriguingly, MeCP2 loss-mediated increases in miR-137, miR-15a, miR-7b and miR-134 each lead to impaired dendritic maturation^{87,89,90}, a major feature of RTT.

Another set of miRNAs are indirectly regulated by MeCP2. miR-199 is robustly upregulated in iPSC-derived neuronal cultures from individuals with RTT who carry a missense mutation encoding a R106W substitution in the MBD of MeCP2, or a frameshift mutation that results in a truncated transcript (V247X), and in MeCP2-deficient mouse neurons⁴¹. Bone morphogenic protein 4 (BMP4) activates SMAD signalling, which in turn promotes the processing of pri-miRNAs, and BMP4 is upregulated in certain MeCP2-mutant models^{91,92}. Therefore, the upregulation of mature miR-199 observed in the RTT cultures probably results from elevated BMP4–SMAD signalling (FIG. 3). The upregulation of miR-199 inhibits extracellular-signal-regulated kinase signalling and disrupts early neuronal development and differentiation⁴¹. Similarly, MeCP2 deficiency results in increased let-7f miRNA expression in the postnatal mouse brain, through reductions in the BDNF-regulated repressor of let-7 miRNA biogenesis, protein lin-28 homologue A (LIN28A)^{93,94}. In turn, this increase in let-7f inhibits the expression of insulin-like growth factor 1 (IGF1) in the postnatal mouse brain⁸⁴. Treatment with the β_2 -adrenergic receptor agonist clenbuterol restores the affected molecular pathway and ameliorates diverse symptoms of *Mecp2*^{-/-} mice and *Mecp2*^{+/-} mice⁸⁴. Taken together, these findings suggest that RTT-relevant mutations⁷⁰ could alter diverse miRNA pathways downstream of MeCP2

and result in disturbances in brain development and maturation.

Last, miRNAs may regulate MeCP2 expression. For example, miR-483-5p regulates MeCP2 isoforms through a human-specific target site in the 3′-untranslated region of *MECP2* in developing neurons⁹⁵. In addition, miR-200a and miR-302c inhibit MeCP2 expression in human embryonic stem cells⁹⁶, and miR-130a regulates neurite and synaptic maturation in the developing cortex through its inhibitory effect on MeCP2 expression⁹⁷. Intriguingly, some miRNAs, such as miR-132 and miR-7b, act in a feedback loop as both targets and regulators of MeCP2 expression to fine-tune MeCP2 levels in the postnatal brain^{85,89,98}.

MeCP2 binds broadly across the genome; therefore, it will remain a challenge to deduce the primary actions of MeCP2 from RNA analyses. Thus, future studies focusing on MeCP2 binding sites (such as methylated DNA) and relating these to miRNA expression will provide crucial information for further analyses of MeCP2 function and about the roles of miRNAs in RTT.

Excitatory and inhibitory circuits

Neurological symptoms in individuals with RTT and in *Mecp2*-mutant mice result from neural circuit alterations brought on by the loss of MeCP2. In *Mecp2*-mutant mice, E/I balance in circuits and brain regions is complexly altered; the maintenance of this balance is crucial for nearly every aspect of normal brain function.

Neuronal circuit deficits with MeCP2 loss are region-specific. Somatosensory cortex neurons in brain slices and cultured neurons from *Mecp2*^{-/-} mice exhibit smaller miniature excitatory postsynaptic currents (mEPSCs), with no changes in miniature inhibitory postsynaptic currents (mIPSCs), resulting in an overall reduced excitation^{29,99,100}. In vivo whole-cell recordings of visually evoked responses in V1 pyramidal neurons reveal a combined reduction of excitatory and inhibitory conductance in *Mecp2*^{-/-} mice, with a net increase in the E/I ratio¹⁰¹. Similarly, there is a shift towards hyperexcitation in the CA1 and CA3 region of the hippocampus in *Mecp2*^{-/-} mice resulting from reduced mIPSCs¹⁰², and synaptic hyperexcitability is observed in brainstem areas, including the locus coeruleus¹⁰³, ventrolateral medulla^{104,105} and nucleus tractus solitarius¹⁰⁶.

As a direct demonstration of the region-specific changes in activity in *Mecp2*^{-/-} mice, expression of the immediate early gene *Fos* is decreased in the forebrain and increased in the hindbrain¹⁰⁶. Gene expression analyses of different neuronal cell types in *Mecp2*-knockout mouse brains showed variation in different sets of genes in these different cell populations⁹⁶. Clinically, selective volume reductions in the dorsal parietal and anterior frontal lobe regions in individuals with RTT also suggest that some brain regions are more affected than others¹⁷.

Overall, these studies highlight an intriguing region specificity of disruption to neural circuits that may explain the clinical features of RTT. For example, reduced excitation and connectivity in forebrain motor areas probably contributes to motor dysfunction in RTT, whereas hyperexcitation in the brainstem

Primary miRNAs

(pri-miRNAs). Large primary precursor, hairpin-structured, microRNA transcripts that are subsequently cleaved to generate intermediate precursor microRNAs and mature microRNAs.

region probably leads to abnormalities in breathing patterns²¹. Similarly, hyperexcitability in the hippocampus probably contributes to the limbic seizures observed in *Mecp2*-mutant mice and people with RTT¹⁰⁷.

Although the mechanisms underlying this region specificity remain unclear, two non-mutually exclusive possibilities can be extrapolated. First, MeCP2 may differentially affect different brain regions. Second, although MeCP2 may function similarly in different regions, some neurons or circuits may be more vulnerable to MeCP2 dysfunction than others, owing to their connectivity (see below).

MeCP2 loss affects excitatory and inhibitory neurons.

Although the molecular effects of MeCP2 seem to be primarily mediated by its MBD and NID (see previous section), the functional effects of MeCP2 are region-specific and cell-type-specific (for example, selective for excitatory or inhibitory neuron subtypes). The functional effects of MeCP2 loss depend on the disruption of E/I imbalance within neuronal circuits¹⁰¹, and neuronal circuits seem to be particularly sensitive to MeCP2 loss from inhibitory neurons²⁴. We therefore suggest that the molecular effects of MeCP2 are not equal across functional cell classes and functional brain regions. Deletion of MeCP2 from functional cell types can provide insights into the primacy of MeCP2 function in specific cell types. One study that used cell-type-specific biotin tagging of MeCP2 in wild-type mice, *Mecp2*^{T158M/y} mice and *Mecp2*^{R106W/y} mice to profile nuclear transcriptomes revealed distinct sets of differentially regulated genes in excitatory and inhibitory neurons in the *Mecp2* mutants⁶⁷. This finding is consistent with the observation that the landscape of DNA methylation varies between excitatory and inhibitory neurons^{61,108}. If the methylation landscape varies across cell types, MeCP2 may exert different molecular actions in different cell types, leading to cell-specific transcriptional alterations with MeCP2 mutations.

Several symptoms of whole-brain *Mecp2* deletion, including ataxia, repetitive behaviours, deficits in motor coordination and obesity, are strikingly recapitulated in mice with *Mecp2* deletion in inhibitory neurons (*Viaat-Mecp2*^{-/-} mice). In addition, the reductions in GABA synthesis and mIPSCs in *Viaat-Mecp2*^{-/-} mice²⁴ are also observed in whole-brain *Mecp2* deletion^{101,109}. Mice lacking MeCP2 in forebrain excitatory neurons exhibit milder behavioural phenotypes, including tremor and anxiety-like behaviours, which are less evident in mice lacking MeCP2 in inhibitory neurons^{110,111}. Furthermore, re-expression of MeCP2 in inhibitory neurons, but not in excitatory neurons, rescues motor coordination deficits in *Mecp2*-knockout mice^{110,112}. These studies indicate that primary deficits in excitatory signalling or inhibitory signalling caused by cell-type-specific MeCP2 loss lead to distinct neurological phenotypes.

Inhibitory neurons and RTT. GABAergic inhibitory neurons account for up to 20% of the total neurons in the brain¹¹³. In particular, subtypes of inhibitory neurons are known to have very different developmental programmes, circuit influences and effects on activity-dependent plasticity^{113,114}. Among the interneuron populations are three major subclasses that directly influence pyramidal

neurons: soma-targeting parvalbumin (PV)-expressing neurons, dendrite-targeting somatostatin (SOM)-expressing neurons and vasoactive intestinal polypeptide (VIP)-expressing neurons^{113,115,116}. SOM⁺ neurons integrate large numbers of inputs¹¹⁷ and seem to be crucial for feedback or top-down modulation¹¹⁸. VIP⁺ neurons are a subset of the serotonin receptor (5-HT_{3A})-expressing subclass of inhibitory neurons and receive top-down inputs; they disinhibit pyramidal neurons by inhibiting other inhibitory neurons^{113,119}. PV⁺ neurons form intricate nests of synaptic contacts on the soma of target pyramidal neurons, providing fast control over their excitability¹¹⁵. PV⁺ neurons, which mainly subserve feedforward inhibition and gain control^{120,121}, have been suggested to be the major drivers of E/I balance in cortical circuits¹²².

Intriguingly, *Mecp2* deletion from different GABAergic neuronal subtypes generates differential and non-overlapping symptoms and phenotypes; mice lacking MeCP2 in PV⁺ neurons show sensory, motor and memory deficits, whereas mice lacking MeCP2 in SOM⁺ neurons exhibit seizures and stereotypies¹²³. These different effects of *Mecp2* deletion in inhibitory neuron subtypes possibly reflect the different developmental and integrative circuit properties of these subtypes.

Anatomically, *Mecp2*^{-/-} mice, particularly juveniles, exhibit more PV⁺ puncta and increased PV⁺ neuron connections in the visual cortex compared with wild-type mice^{109,124}. In addition, *Mecp2*^{-/-} mice show alterations in NMDAR subunit expression, with a greater NR2A:NR2B ratio in PV⁺ neurons, suggesting abnormally accelerated NMDAR maturation¹²⁵. Reducing NR2A expression in *Mecp2*^{-/-} mice (achieved by crossing heterozygous *Mecp2*^{+/-} mice with NR2A-deficient males) reduces the PV⁺-cell hyperconnectivity and prevents the regression of visual acuity^{109,125}. However, reductions in inhibition — including reduced PV⁺ neuron responses — have consistently been reported in V1 of adult *Mecp2*^{-/-} mice, *Viaat-Mecp2*^{-/-} mice (which lack MeCP2 from all GABAergic cells) and PV⁺ cell-specific *Mecp2*^{-/-} mice^{24,101}. In addition, brain-wide *Mecp2*^{-/-} mice and mice lacking *Mecp2* from PV⁺ cells display impaired processing of complex visual stimuli by neurons in the visual cortex, consistent with severely reduced inhibition in vivo^{101,126}.

One proposed explanation to reconcile reports of hyperconnectivity of PV⁺ neurons with reduced functional inhibition of pyramidal neurons in *Mecp2*-mutant mice is that the effectiveness of GABA as an inhibitory neurotransmitter may be altered in RTT mouse models. Indeed, recent studies (see below) demonstrate that GABA is less hyperpolarizing in *Mecp2*-mutant mice¹⁰¹ and in iPSC-derived neurons generated from RTT patients¹²⁷. Thus, PV⁺ neuron hyperconnectivity may represent a homeostatic, compensatory response to the reduction in inhibition in cortical circuits (FIG. 4). Neurodevelopmental disorders are marked by germline or early genetic mutations and a cascade of changes that are superimposed on developmental programmes of growth and plasticity; altogether, these factors lead to altered neuronal wiring and function. Thus, direct and indirect effects are deeply intertwined in creating functional as well as dysfunctional neural circuits³³.

Stereotypies

Diagnostic repetitive behaviours. Patients with RTT develop repetitive hand movements, a diagnostic criterion for RTT. Mouse models of RTT also develop repetitive behaviour.

KCC2:NKCC1 imbalance in RTT. The neuronal cation–chloride co-transporters KCC2 (a K^+/Cl^- exporter) and NKCC1 (a $Na^+/K^+/Cl^-$ importer) set the transmembrane chloride gradient and determine the maturation of GABAergic inhibition^{128,129}. During typical brain development, GABAergic currents shift their polarity from depolarization to hyperpolarization as the KCC2:NKCC1 ratio increases. GABA typically hyperpolarizes (inhibits) mature neurons, as they have very low intracellular chloride levels, whereas GABA has less inhibitory, or even depolarizing (excitatory), effects on immature neurons, which show high intracellular chloride concentrations¹³⁰. Increased intracellular chloride concentrations and KCC2–NKCC1 imbalance have been reported in animal models of fragile X syndrome and in other neurological and neuropsychiatric disorders^{131,132}.

Recent evidence suggests that such alterations are also present in RTT. Cerebrospinal fluid samples from individuals with RTT have revealed that levels of KCC2 relative to NKCC1 are reduced¹³³. The expression of KCC2 in iPSC-derived neuronal cultures generated from individuals with RTT, and in V1 of *Mecp2*^{-/-} mice, is also reduced, and this reduction causes the reversal potential of GABA to be less negative than in controls^{101,127}. Overexpression of KCC2 in iPSC-derived neurons generated from individuals with RTT rescues this functional deficit¹²⁷. IGF1 treatment, which partially rescues behavioural and synaptic deficits in mutant mouse models of RTT^{29,134}, also increases the KCC2:NKCC1 ratio by promoting KCC2 expression and restoring the inhibitory action of GABA¹³⁵. Thus, the circuit effects of MeCP2 loss crucially include reduced inhibition owing to reduced KCC2 expression and a reduced KCC2:NKCC1 ratio (FIG. 4).

Activity-dependent plasticity

Neuronal circuits are subject to experience-driven^{136,137} or learning-driven^{138–140} changes that cause neurons within a microcircuit or an ensemble to modify their functional connectivity and responses¹⁴¹. The development of excitatory¹⁴² and inhibitory¹⁴³ microcircuits and the maintenance of E/I balance¹²² require activity-dependent mechanisms, including Hebbian and homeostatic plasticity. Whereas the term Hebbian plasticity describes associative and input-specific potentiation or depression of the strength of a synapse, homeostatic plasticity describes a negative feedback-driven mechanism that maintains appropriate levels of overall activity and synaptic strength in neurons and circuits¹⁴⁴. Together, these mechanisms enable neuronal activity to induce pleiotropic effects on gene expression to regulate synaptic molecule levels and function, and allow neuronal circuits to respond dynamically to experience.

Impaired homeostatic machinery resulting in altered experience-dependent refinement of circuits may be a core functional mechanism underlying autism and RTT^{33,145,146}. Recent evidence indicates that early-life experience or changes in activity, such as that induced by dark rearing, alter methylation, which influences MeCP2 binding and hence gene expression⁶¹. The reverse is also true: *Mecp2* mutations result in changes

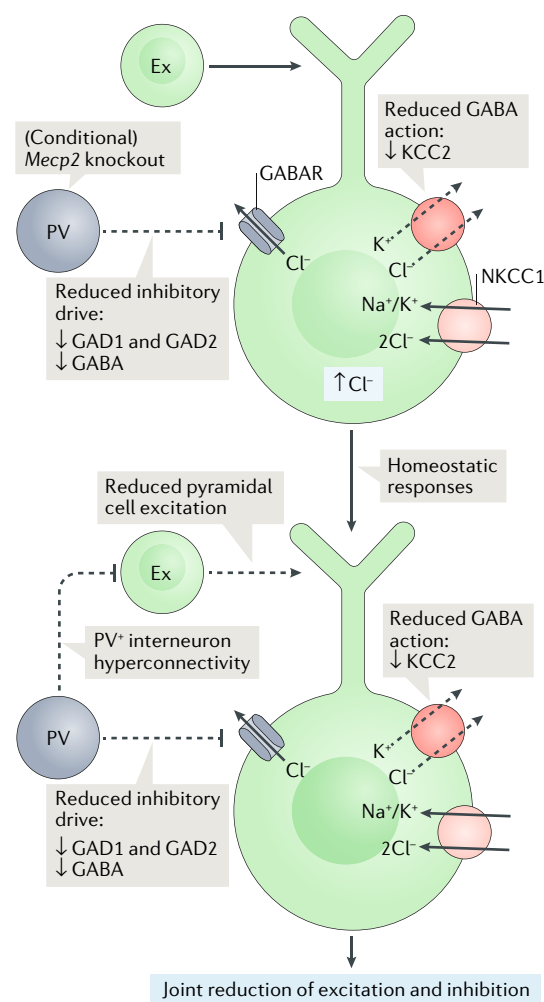


Fig. 4 | Effects of MeCP2 deficiency in inhibitory neurons and excitation–inhibition balance. Deletion of methyl-CpG-binding protein 2 (MeCP2) in all forebrain neurons or only in inhibitory neurons (such as parvalbumin (PV)-positive neurons) reduces GABA synthesis, glutamate decarboxylase 1 (GAD1) and GAD2 levels and miniature inhibitory postsynaptic currents, and thus reduces inhibition onto excitatory pyramidal neurons (green cells). The effectiveness of GABA as an inhibitory neurotransmitter is also altered in MeCP2-deficient neurons. Reduced levels of the K^+/Cl^- exporter KCC2 relative to the $Na^+/K^+/Cl^-$ symporter NKCC1 have been found in cells with mutations in the gene encoding MeCP2, resulting in neurons with higher levels of intracellular chloride, and hence a reduction in the reversal potential for GABA and reduced inhibitory drive to pyramidal neurons. We suggest that homeostatic compensation for the reduction in inhibition leads to hyperconnectivity of PV⁺ neurons and reduced excitatory drive onto pyramidal neurons, resulting in a joint reduction of excitation and inhibition in cortical circuits. Dashed arrows depict reduced function and solid arrows depict normal function. Ex, excitatory; GABAR, GABA receptor.

in experience-dependent plasticity, including (but not limited to) homeostatic plasticity^{147–149}. For example, retinogeniculate synapses in *Mecp2*-null mice show a lack of experience-dependent refinement, despite an apparently normal initial development of synaptic

Fragile X syndrome

A genetic disorder, caused by mutations in the *FMR1* gene, that includes developmental problems such as learning disabilities and cognitive impairment.

Reversal potential

The membrane potential at which a given ion has no net current flow through the cell membrane.

Retinogeniculate synapses

Synaptic connections projecting directly from retinal ganglion cells in the retina to the neurons in the lateral geniculate nucleus.

connectivity¹⁵⁰. *Mecp2*-mutant mice also show substantial impairments in experience-dependent plasticity in the visual cortex^{29,134}. Closure of one eye during a critical period of development can cause ocular dominance plasticity, which leads to decreased and increased visual cortex responses to the deprived and non-deprived eye, respectively¹⁵¹. In *Mecp2*-mutant mice, the critical period during which ocular dominance plasticity can be induced is prolonged and extends into adulthood, mainly because adult mice still show a persistent homeostatic upscaling of cortical responses to stimulation of the non-deprived eye¹³⁴. Similarly, PV⁺ cell-specific *Mecp2*^{-/-} mice exhibit deprivation-induced plasticity in adulthood, accompanied by a reduction of inhibition¹⁰¹. This persistent experience-dependent plasticity is corrected by treatment with IGF1, which has crucial roles in the maturation of PV⁺ circuits as well as GABAergic inhibition^{29,101}. A critical level of inhibition is also required to initiate plasticity. PV⁺-cell-specific *Mecp2*^{-/-} mice fail to show ocular dominance plasticity during the critical period, consistent with a reduction of inhibition, and restoring inhibition through intracortical infusion of the GABA agonist diazepam restores plasticity¹²⁶.

The activity-induced phosphorylation of MeCP2 at the NID is important for activity-dependent plasticity. Neuronal activity induces phosphorylation of MeCP2 at several sites, including T308 (REF.⁷¹). Phosphorylation at T308 blocks the interaction of MeCP2 with the NCoR-SMRT complex, thus suppressing the ability of MeCP2 to repress transcription⁷¹. A human RTT-associated missense mutation in *MECP2*, leading to a R306C substitution, abolishes phosphorylation of MeCP2 at T308 and the interaction of MeCP2 with the NCoR-SMRT complex^{68,71}. Importantly, both *Mecp2*^{R306C} and *Mecp2*^{T308A} knock-in mice show RTT-like behavioural phenotypes, although *Mecp2*^{R306C} knock-in mice display a more severe phenotype^{68,71}. A subset of activity-regulated genes that influence inhibitory neurons, including *Npas4* and *Bdnf*, are downregulated in *Mecp2*^{T308A} mice⁷¹, possibly providing a molecular mechanism for the alterations in inhibition in these animals. Thus, dysregulation of activity-dependent signalling that is normally mediated by the NID of MeCP2 likely contributes crucially to RTT pathophysiology.

Conclusions and perspectives

MeCP2 has spatiotemporally diverse effects depending on brain regions, phases of development and cell types. MeCP2 regulates multiple stages of brain development, including prenatal neurogenesis, postnatal development of synaptic connections and function, and experience-dependent refinement of neural circuits. Like many other brain disorders, RTT is associated with dysfunction reflective of functional failure at several levels, including the levels of gene regulation, molecular expression, synaptic function and neuronal circuitry, and at various developmental stages.

Accumulating evidence indicates that the primary role of MeCP2 is transcriptional repression, with the MBD and NID of MeCP2 connecting methylated DNA with the NCoR-SMRT co-repressor complex. However,

it remains unclear how certain missense mutations, such as that encoding the T158M substitution in the MBD, confer more severe clinical symptoms than do mutations affecting the NID (such as that encoding the R306C substitution)¹⁵². Examining transcriptomes associated with different RTT mutations could provide critical insights into the differences in clinical severity.

At the molecular level, one major question is how MeCP2 loss produces the subtle yet widespread effects on neuron-enriched long genes, key activity-dependent signalling pathways and a large array of miRNAs. All of these effects may reflect the ability of MeCP2 to interact widely across the genome. To better understand the functions of MeCP2, an important next step will be to evaluate, in detail, the molecular events that occur following MeCP2 binding to the gene bodies across the genome. Another feature of RTT to be resolved is a reduction in total RNA^{57,74}, which is potentially due to suboptimal functioning of transcriptional machinery and which may explain the large number of downregulated genes in RTT.

The molecular mechanisms of MeCP2 loss manifest as dysfunction in brain circuits. The wiring and function of brain circuits depend on proper functioning of inhibitory and excitatory neurons and maintenance of E/I balance, all of which require MeCP2. A major functional contributor to RTT phenotypes is alteration of inhibition, owing to altered gene expression in GABAergic neurons and a reduction in inhibitory drive onto pyramidal cells. Importantly, the NID mediates activity-dependent functions of MeCP2 and hence probably influences activity-dependent maturation of inhibitory neurons and the function of GABA receptors.

A key outstanding question is how a loss of the molecular functions of MeCP2 leads to the functional circuit deficits in RTT phenotypes. Are there cell-type-specific effects of the loss of the normal molecular mechanisms of MeCP2? In particular, it will be important to establish the effects of mutations affecting the MBD or NID on PV⁺ neurons and to investigate how these domains shape the properties of PV⁺ neurons and hence functional circuits. Experimentally, combining single-cell RNA sequencing and physiology offers an opportunity to dissect such alterations, thus bridging molecular dysregulation and circuit dysfunction following MeCP2 loss.

The development of treatment strategies (BOX 3) must take into account the functional properties of MeCP2 discussed in this Review and consider the therapeutic target, brain region, cell type and time of application. More directly targeted therapeutics that are administered as early as possible are likely to be the most effective. Gene therapy is a potentially powerful approach (BOX 3), although, notably, the protein level of MeCP2 must be tightly titrated, as overdosage of MeCP2, similar to *MECP2* duplication, would lead to severe neurological consequences^{153–155}. A fuller understanding of RTT based on our increasing knowledge of the neurobiology and function of MeCP2, together with rapidly advancing genome-editing tools, could lead to an effective treatment for RTT.

Published online: 08 May 2018

1. Chahrouh, M. & Zoghbi, H. Y. The story of Rett syndrome: from clinic to neurobiology. *Neuron* **56**, 422–437 (2007).
2. Laurvick, C. L. et al. Rett syndrome in Australia: a review of the epidemiology. *J. Pediatr.* **148**, 347–352 (2006).
3. Trappe, R. et al. *MECP2* mutations in sporadic cases of Rett syndrome are almost exclusively of paternal origin. *Am. J. Hum. Genet.* **68**, 1093–1101 (2001).
4. Neul, J. L. et al. Specific mutations in *methyl-CpG-binding protein 2* confer different severity in Rett syndrome. *Neurology* **70**, 1313–1321 (2008).
5. Neul, J. L. et al. Rett syndrome: revised diagnostic criteria and nomenclature. *Ann. Neurol.* **68**, 944–950 (2010).
6. Schüle, B., Armstrong, D. D., Vogel, H., Oviedo, A. & Francke, U. Severe congenital encephalopathy caused by *MECP2* null mutations in males: central hypoxia and reduced neuronal dendritic structure. *Clin. Genet.* **74**, 116–126 (2008).
7. Lewis, J. D. et al. Purification, sequence, and cellular localization of a novel chromosomal protein that binds to methylated DNA. *Cell* **69**, 905–914 (1992).
8. Chahrouh, M. et al. *MeCP2*, a key contributor to neurological disease, activates and represses transcription. *Science* **320**, 1224–1229 (2008).
9. Cheng, T. L. et al. *MeCP2* suppresses nuclear microRNA processing and dendritic growth by regulating the DGCR8/Drosha complex. *Dev. Cell* **28**, 547–560 (2014).
10. Dolce, A. et al. Rett syndrome and epilepsy: an update for child neurologists. *Pediatr. Neurol.* **48**, 337–345 (2013).
11. Leonard, H., Cobb, S. & Downs, J. Clinical and biological progress over 50 years in Rett syndrome. *Nat. Rev. Neurol.* **13**, 37–51 (2016).
12. von Tetzchner, S. et al. Vision, cognition and developmental characteristics of girls and women with Rett syndrome. *Dev. Med. Child Neurol.* **38**, 212–225 (1996).
13. LeBlanc, J. J. et al. Visual evoked potentials detect cortical processing deficits in Rett syndrome. *Ann. Neurol.* **78**, 775–786 (2015).
14. Neul, J. L. The relationship of Rett syndrome and *MECP2* disorders to autism. *Dialogues Clin. Neurosci.* **14**, 253–262 (2012).
15. American Psychiatric Association (eds). Diagnostic and Statistical Manual of Mental Disorders, Fifth Edition (American Psychiatric Publishing, 2013).
16. Armstrong, D., Dunn, J. K., Antalfy, B. & Trivedi, R. Selective dendritic alterations in the cortex of Rett syndrome. *J. Neuropathol. Exp. Neurol.* **54**, 195–201 (1995).
17. Carter, J. C. et al. Selective cerebral volume reduction in Rett Syndrome: a multiple-approach MR imaging study. *AJNR Am. J. Neuroradiol.* **29**, 436–441 (2008).
18. Amir, R. E. et al. Rett syndrome is caused by mutations in X-linked *MECP2*, encoding methyl-CpG-binding protein 2. *Nat. Genet.* **23**, 185–188 (1999).
19. Chen, R. Z., Akbarian, S., Tudor, M. & Jaenisch, R. Deficiency of methyl-CpG binding protein-2 in CNS neurons results in a Rett-like phenotype in mice. *Nat. Genet.* **27**, 327–331 (2001).
20. Guy, J., Hendrich, B., Holmes, M., Martin, J. E. & Bird, A. A mouse *Mecp2*-null mutation causes neurological symptoms that mimic Rett syndrome. *Nat. Genet.* **27**, 322–326 (2001).
21. Ward, C. S. et al. *MeCP2* is critical within HoxB1-derived tissues of mice for normal lifespan. *J. Neurosci.* **31**, 10359–10370 (2011).
22. Huang, T.-W. et al. Progressive changes in a distributed neural circuit underlie breathing abnormalities in mice lacking *MeCP2*. *J. Neurosci.* **36**, 5572–5586 (2016).
23. Samaco, R. C. et al. Loss of *MeCP2* in aminergic neurons causes cell-autonomous defects in neurotransmitter synthesis and specific behavioral abnormalities. *Proc. Natl Acad. Sci. USA* **106**, 21966–21971 (2009).
24. Chao, H.-T. et al. Dysfunction in GABA signalling mediates autism-like stereotypies and Rett syndrome phenotypes. *Nature* **468**, 263–269 (2010).
This study shows that mice with *Mecp2* deleted only in GABAergic neurons recapitulate many of the deficits observed in global knockout mice, suggesting that *MeCP2* is crucial for the normal function of GABAergic neurons and that the dysfunction of GABAergic neurons contributes to RTT phenotypes.
25. Lioy, D. T. et al. A role for glia in the progression of Rett's syndrome. *Nature* **475**, 497–500 (2011). **This paper shows that expression of *MeCP2* exclusively in astrocytes rescues some of the major deficits in *MeCP2*-deficient mice, suggesting that *MeCP2* does not function only in neurons and has non-cell-autonomous effects.**
26. Derecki, N. C. et al. Wild-type microglia arrest pathology in a mouse model of Rett syndrome. *Nature* **484**, 105–109 (2012).
27. Wang, J. et al. Wild-type microglia do not reverse pathology in mouse models of Rett syndrome. *Nature* **521**, E1–E4 (2015).
28. Schafer, D. P. et al. Microglia contribute to circuit defects in *Mecp2* null mice independent of microglia-specific loss of *Mecp2* expression. *eLife* **5**, e15224 (2016).
29. Tropea, D. et al. Partial reversal of Rett Syndrome-like symptoms in *MeCP2* mutant mice. *Proc. Natl Acad. Sci. USA* **106**, 2029–2034 (2009).
This paper shows that systemic treatment of *MeCP2*-mutant mice with a truncated form of IGF1 restores excitatory synapses and improves neurological deficits.
30. Fukuda, T., Itoh, M., Ichikawa, T., Washiyama, K. & Goto, Y. Delayed maturation of neuronal architecture and synaptogenesis in cerebral cortex of *Mecp2*-deficient mice. *J. Neuropathol. Exp. Neurol.* **64**, 537–544 (2005).
31. Smrt, R. D. et al. *Mecp2* deficiency leads to delayed maturation and altered gene expression in hippocampal neurons. *Neurobiol. Dis.* **27**, 77–89 (2007).
32. Cohen, S. & Greenberg, M. E. Communication between the synapse and the nucleus in neuronal development, plasticity, and disease. *Annu. Rev. Cell Dev. Biol.* **24**, 183–209 (2008).
33. Nelson, S. B. & Valakh, V. Excitatory/inhibitory balance and circuit homeostasis in autism spectrum disorders. *Neuron* **87**, 684–698 (2015).
34. Shahbazian, M. D., Antalfy, B., Armstrong, D. L. & Zoghbi, H. Y. Insight into Rett syndrome: *MeCP2* levels display tissue- and cell-specific differences and correlate with neuronal maturation. *Hum. Mol. Genet.* **11**, 115–124 (2002).
35. Kishi, N. & Macklis, J. D. *MECP2* is progressively expressed in post-migratory neurons and is involved in neuronal maturation rather than cell fate decisions. *Mol. Cell Neurosci.* **27**, 306–321 (2004).
36. Fehr, S. et al. Altered attainment of developmental milestones influences the age of diagnosis of Rett syndrome. *J. Child Neurol.* **26**, 980–987 (2011).
37. Einspieler, C., Kerr, A. M. & Prechtl, H. F. R. Is the early development of girls with Rett disorder really normal? *Pediatr. Res.* **57**, 696–700 (2005).
38. Einspieler, C., Kerr, A. M. & Prechtl, H. F. R. Abnormal general movements in girls with Rett disorder: the first four months of life. *Brain Dev.* **27**, S8–S13 (2005).
39. Marschik, P. B. et al. Changing the perspective on early development of Rett syndrome. *Res. Dev. Disabil.* **34**, 1236–1239 (2013).
40. Neul, J. L. et al. Developmental delay in Rett syndrome: data from the natural history study. *J. Neurodev. Disord.* **6**, 20 (2014).
41. Mellios, N. et al. *MeCP2*-regulated miRNAs control early human neurogenesis through differential effects on ERK and AKT signaling. *Mol. Psychiatry* **23**, 1051–1065 (2018).
This study examines cerebral organoids of iPSC-derived cells from individuals with RTT, showing dysregulation of miR-199 and miR-214 signalling and early developmental defects.
42. Bedogni, F. et al. Defects during *Mecp2* null embryonic cortex development precede the onset of overt neurological symptoms. *Cereb. Cortex* **26**, 2517–2529 (2016).
43. Kim, K. Y., Hysolli, E. & Park, I. H. Neuronal maturation defect in induced pluripotent stem cells from patients with Rett syndrome. *Proc. Natl Acad. Sci. USA* **108**, 14169–14174 (2011).
44. Tsujimura, K., Abematsu, M., Kohyama, J., Namihiara, M. & Nakashima, K. Neuronal differentiation of neural precursor cells is promoted by the methyl-CpG-binding protein *MeCP2*. *Exp. Neurol.* **219**, 104–111 (2009).
45. Stancheva, I. et al. A mutant form of *MeCP2* protein associated with human Rett syndrome cannot be displaced from methylated DNA by notch in *Xenopus* embryos. *Mol. Cell* **12**, 425–435 (2003).
46. Gao, H. et al. *Mecp2* regulates neural cell differentiation by suppressing the *Id1* to *Her2* axis in zebrafish. *J. Cell Sci.* **128**, 2340–2350 (2015).
47. Guy, J., Gan, J., Selfridge, J., Cobb, S. & Bird, A. Reversal of neurological defects in a mouse model of Rett syndrome. *Science* **315**, 1143–1147 (2007).
48. Giacometti, E., Luikenhuis, S., Beard, C. & Jaenisch, R. Partial rescue of *MeCP2* deficiency by postnatal activation of *MeCP2*. *Proc. Natl Acad. Sci. USA* **104**, 1931–1936 (2007).
49. McGraw, C. M., Samaco, R. C. & Zoghbi, H. Y. Adult neural function requires *MeCP2*. *Science* **333**, 186 (2011).
50. Nguyen, M. V. C. et al. *MeCP2* is critical for maintaining mature neuronal networks and global brain anatomy during late stages of postnatal brain development and in the mature adult brain. *J. Neurosci.* **32**, 10021–10034 (2012).
51. Du, F. et al. Acute and crucial requirement for *MeCP2* function upon transition from early to late adult stages of brain maturation. *Hum. Mol. Genet.* **25**, 1690–1702 (2016).
52. Swiech, L. et al. In vivo interrogation of gene function in the mammalian brain using CRISPR-Cas9. *Nat. Biotechnol.* **33**, 102–106 (2014).
53. Lyst, M. J. & Bird, A. Rett syndrome: a complex disorder with simple roots. *Nat. Rev. Genet.* **16**, 261–275 (2015).
54. Meehan, R., Lewis, J. D. & Bird, A. P. Characterization of *MECP2*, a vertebrate DNA binding protein with affinity for methylated DNA. *Nucleic Acids Res.* **20**, 5085–5092 (1992).
55. Nan, X. et al. Transcriptional repression by the methyl-CpG-binding protein *MeCP2* involves a histone deacetylase complex. *Nature* **393**, 386–389 (1998).
56. Kinde, B., Gabel, H. W., Gilbert, C. S., Griffith, E. C. & Greenberg, M. E. Reading the unique DNA methylation landscape of the brain: non-CpG methylation, hydroxymethylation, and *MeCP2*. *Proc. Natl Acad. Sci. USA* **112**, 6800–6806 (2015).
57. Lager, S. et al. *MeCP2* recognizes cytosine methylated tri-nucleotide and di-nucleotide sequences to tune transcription in the mammalian brain. *PLoS Genet.* **13**, e1006793 (2017).
58. Guo, J. U. et al. Distribution, recognition and regulation of non-CpG methylation in the adult mammalian brain. *Nat. Neurosci.* **17**, 215–222 (2013).
59. Gabel, H. W. et al. Disruption of DNA-methylation-dependent long gene repression in Rett syndrome. *Nature* **522**, 89–93 (2015).
This paper demonstrates that *MeCP2* binds to mCA sites and this binding represses the expression of long genes.
60. Kinde, B., Wu, D. Y., Greenberg, M. E. & Gabel, H. W. DNA methylation in the gene body influences *MeCP2*-mediated gene repression. *Proc. Natl Acad. Sci. USA* **113**, 15114–15119 (2016).
61. Stroud, H. et al. Early-life gene expression in neurons modulates lasting epigenetic states. *Cell* **171**, 1151–1164.e16 (2017).
This study shows that DNMT3A specifies the pattern of DNA methylation at CA sites, which in turn directs the binding of *MeCP2* and regulates the transcription of genes.
62. Mellén, M., Ayata, P., Dewell, S., Kraucionis, S. & Heintz, N. *MeCP2* binds to 5hmC enriched within active genes and accessible chromatin in the nervous system. *Cell* **151**, 1417–1430 (2012).
63. Mellén, M., Ayata, P. & Heintz, N. 5-Hydroxymethylcytosine accumulation in postmitotic neurons results in functional demethylation of expressed genes. *Proc. Natl Acad. Sci. USA* **114**, E7812–E7821 (2017).
64. Skene, P. J. et al. Neuronal *MeCP2* is expressed at near histone-octamer levels and globally alters the chromatin state. *Mol. Cell* **37**, 457–468 (2010).
65. Shah, R. R. & Bird, A. P. *MeCP2* mutations: progress towards understanding and treating Rett syndrome. *Genome Med.* **9**, 17 (2017).
66. Sugino, K. et al. Cell-type-specific repression by methyl-CpG-binding protein 2 is biased toward long genes. *J. Neurosci.* **34**, 12877–12883 (2014).
67. Johnson, B. S. et al. Biotin tagging of *MeCP2* in mice reveals contextual insights into the Rett syndrome transcriptome. *Nat. Med.* **23**, 1203–1214 (2017).
68. Lyst, M. J. et al. Rett syndrome mutations abolish the interaction of *MeCP2* with the NCoR/SMRT co-repressor. *Nat. Neurosci.* **16**, 898–902 (2013).
This study identifies the NID in *MeCP2* and demonstrates that RTT-causing mutations in the NID abolish the interaction between *MeCP2* and the NCoR-SMRT co-repressor complexes.
69. Kruusvee, V. et al. Structure of the *MeCP2*-TBLR1 complex reveals a molecular basis for Rett syndrome and related disorders. *Proc. Natl Acad. Sci. USA* **114**, E3243–E3250 (2017).
70. Tillotson, R. et al. Radically truncated *MeCP2* rescues Rett syndrome-like neurological defects. *Nature* **550**, 398–401 (2017).

This study establishes that truncated MeCP2 protein retaining only the MBD and NID is able to rescue neurological symptoms when introduced into MeCP2-deficient mice.

71. Ebert, D. H. et al. Activity-dependent phosphorylation of MECP2 T308 regulates interaction with NCoR. *Nature* **499**, 341–345 (2013).
72. Ben-Shachar, S., Chahrour, M., Thaller, C., Shaw, C. A. & Zoghbi, H. Y. Mouse models of MeCP2 disorders share gene expression changes in the cerebellum and hypothalamus. *Hum. Mol. Genet.* **18**, 2431–2442 (2009).
73. Tudor, M., Akbarian, S., Chen, R. Z. & Jaenisch, R. Transcriptional profiling of a mouse model for Rett syndrome reveals subtle transcriptional changes in the brain. *Proc. Natl Acad. Sci. USA* **99**, 15536–15541 (2002).
74. Li, Y. et al. Global transcriptional and translational repression in human-embryonic-stem-cell-derived Rett syndrome neurons. *Cell Stem Cell* **13**, 446–458 (2013).
- This study inactivates MECP2 in human iPSC-derived neurons and shows a global alteration of RNA transcripts.**
75. Nott, A. et al. Histone deacetylase 3 associates with MeCP2 to regulate FOXO and social behavior. *Nat. Neurosci.* **19**, 1497–1505 (2016).
76. Ciernia, A. V. & LaSalle, J. The landscape of DNA methylation amid a perfect storm of autism aetiologies. *Nat. Rev. Neurosci.* **17**, 411–423 (2016).
77. Baker, S. A. et al. An AT-hook domain in MeCP2 determines the clinical course of Rett syndrome and related disorders. *Cell* **152**, 984–996 (2013).
78. Heckman, L. D., Chahrour, M. H. & Zoghbi, H. Y. Rett-causing mutations reveal two domains critical for MeCP2 function and for toxicity in MECP2 duplication syndrome mice. *eLife* **3**, e02676 (2014).
79. Nan, X. et al. Interaction between chromatin proteins MECP2 and ATRX is disrupted by mutations that cause inherited mental retardation. *Proc. Natl Acad. Sci. USA* **104**, 2709–2714 (2007).
80. Krol, J., Loedige, I. & Filipowicz, W. The widespread regulation of microRNA biogenesis, function and decay. *Nat. Rev. Genet.* **11**, 597–610 (2010).
81. Mellios, N. & Sur, M. The emerging role of microRNAs in schizophrenia and autism spectrum disorders. *Front. Psychiatry* **3**, 39 (2012).
82. Wu, H. et al. Genome-wide analysis reveals methyl-CpG-binding protein 2-dependent regulation of microRNAs in a mouse model of Rett syndrome. *Proc. Natl Acad. Sci. USA* **107**, 18161–18166 (2010).
83. Urdinguio, R. G. et al. Disrupted microRNA expression caused by Mecp2 loss in a mouse model of Rett syndrome. *Epigenetics* **5**, 656–663 (2010).
84. Mellios, N. et al. β_2 -Adrenergic receptor agonist ameliorates phenotypes and corrects microRNA-mediated IGF1 deficits in a mouse model of Rett syndrome. *Proc. Natl Acad. Sci. USA* **111**, 9947–9952 (2014).
85. Klein, M. E. et al. Homeostatic regulation of MeCP2 expression by a CREB-induced microRNA. *Nat. Neurosci.* **10**, 1513–1514 (2007).
86. Nomura, T. et al. MeCP2-dependent repression of an imprinted miR-184 released by depolarization. *Hum. Mol. Genet.* **17**, 1192–1199 (2008).
87. Gao, Y. et al. Inhibition of miR-15a promotes BDNF expression and rescues dendritic maturation deficits in MeCP2-deficient neurons. *Stem Cells* **33**, 1618–1629 (2015).
88. Szulwach, K. E. et al. Cross talk between microRNA and epigenetic regulation in adult neurogenesis. *J. Cell Biol.* **189**, 127–141 (2010).
89. Chen, Y., Shin, B. C., Thamotharan, S. & Devaskar, S. U. Differential methylation of the microRNA 7b gene targets postnatal maturation of murine neuronal *Mecp2* gene expression. *Dev. Neurobiol.* **74**, 407–425 (2014).
90. Smrt, R. D. et al. MicroRNA miR-137 regulates neuronal maturation by targeting ubiquitin ligase mind bomb-1. *Stem Cells* **28**, 1060–1070 (2010).
91. Davis, B. N., Hilyard, A. C., Lagna, G. & Hata, A. SMAD proteins control DROSHA-mediated microRNA maturation. *Nature* **454**, 56–61 (2008).
92. Li, H., Zhong, X., Chau, K. F., Williams, E. C. & Chang, Q. Loss of activity-induced phosphorylation of MeCP2 enhances synaptogenesis, LTP and spatial memory. *Nat. Neurosci.* **14**, 1001–1008 (2011).
93. Huang, Y.-W. A., Ruiz, C. R., Eyster, E. C. H., Lin, K. & Meffert, M. K. Dual regulation of microRNA biogenesis generates target specificity in neurotrophin-induced protein synthesis. *Cell* **148**, 933–946 (2012).
94. Piskounova, E. et al. Lin28A and Lin28B inhibit let-7 microRNA biogenesis by distinct mechanisms. *Cell* **147**, 1066–1079 (2011).
95. Han, K. et al. Human-specific regulation of MeCP2 levels in fetal brains by microRNA miR-483-5p. *Genes Dev.* **27**, 485–490 (2013).
96. Rodrigues, D. C. et al. MeCP2 is post-transcriptionally regulated during human neurodevelopment by combinatorial action of RNA-binding proteins and miRNAs. *Cell Rep.* **17**, 720–734 (2016).
97. Zhang, Y. et al. MiR-130a regulates neurite outgrowth and dendritic spine density by targeting MeCP2. *Protein Cell* **7**, 489–500 (2016).
98. Im, H. I., Hollander, J. A., Bali, P. & Kenny, P. J. MeCP2 controls BDNF expression and cocaine intake through homeostatic interactions with microRNA-212. *Nat. Neurosci.* **13**, 1120–1127 (2010).
99. Dani, V. S. et al. Reduced cortical activity due to a shift in the balance between excitation and inhibition in a mouse model of Rett syndrome. *Proc. Natl Acad. Sci. USA* **102**, 12560–12565 (2005).
100. Nelson, E. D., Kavalali, E. T. & Monteggia, L. M. MeCP2-dependent transcriptional repression regulates excitatory neurotransmission. *Curr. Biol.* **16**, 710–716 (2006).
101. Banerjee, A. et al. Jointly reduced inhibition and excitation underlies circuit-wide changes in cortical processing in Rett syndrome. *Proc. Natl Acad. Sci. USA* **113**, E7287–E7296 (2016). **This study shows that visually driven excitatory and inhibitory conductances are both reduced in MeCP2-deficient mice in vivo, and that PV⁺-cell-specific MeCP2 deletion recapitulates effects of global MeCP2 deletion on cortical circuits.**
102. Calfa, G., Li, W., Rutherford, J. M. & Pozzo-Miller, L. Excitation/inhibition imbalance and impaired synaptic inhibition in hippocampal area CA3 of *Mecp2* knockout mice. *Hippocampus* **25**, 159–168 (2015).
103. Taneja, P. et al. Pathophysiology of locus ceruleus neurons in a mouse model of Rett syndrome. *J. Neurosci.* **29**, 12187–12195 (2009).
104. Abdala, A. P. L., Dutschmann, M., Bissonnette, J. M. & Paton, J. F. R. Correction of respiratory disorders in a mouse model of Rett syndrome. *Proc. Natl Acad. Sci. USA* **107**, 18208–18213 (2010).
105. Medrihan, L. et al. Early defects of GABAergic synapses in the brain stem of a MeCP2 mouse model of Rett syndrome. *J. Neurophysiol.* **99**, 112–121 (2008).
106. Kron, M. et al. Brain activity mapping in *Mecp2* mutant mice reveals functional deficits in forebrain circuits, including key nodes in the default mode network, that are reversed with ketamine treatment. *J. Neurosci.* **32**, 13860–13872 (2012).
107. Calfa, G. et al. Network hyperexcitability in hippocampal slices from *Mecp2* mutant mice revealed by voltage-sensitive dye imaging. *J. Neurophysiol.* **105**, 1768–1784 (2011).
108. Mo, A. et al. Epigenomic signatures of neuronal diversity in the mammalian brain. *Neuron* **86**, 1369–1384 (2015).
109. Durand, S. et al. NMDA receptor regulation prevents regression of visual cortical function in the absence of *Mecp2*. *Neuron* **76**, 1078–1090 (2012).
110. Meng, X. et al. Manipulations of MeCP2 in glutamatergic neurons highlight their contributions to Rett and other neurological disorders. *eLife* **5**, e14199 (2016).
111. Gemelli, T. et al. Postnatal loss of methyl-CpG binding protein 2 in the forebrain is sufficient to mediate behavioral aspects of Rett syndrome in mice. *Biol. Psychiatry* **59**, 468–476 (2006).
112. Ure, K. et al. Restoration of *Mecp2* expression in GABAergic neurons is sufficient to rescue multiple disease features in a mouse model of Rett syndrome. *eLife* **5**, e14198 (2016).
113. Rudy, B., Fishell, G., Lee, S. H. & Hjerling-Leffler, J. Three groups of interneurons account for nearly 100% of neocortical GABAergic neurons. *Dev. Neurobiol.* **71**, 45–61 (2011).
114. Kpeces, A. & Fishell, G. Interneuron cell types are fit to function. *Nature* **505**, 318–326 (2014).
115. Fishell, G. & Rudy, B. Mechanisms of inhibition within the telencephalon: 'where the wild things are'. *Annu. Rev. Neurosci.* **34**, 535–567 (2011).
116. Pfeffer, C. K., Xue, M., He, M., Huang, Z. J. & Scanziani, M. Inhibition of inhibition in visual cortex: the logic of connections between molecularly distinct interneurons. *Nat. Neurosci.* **16**, 1068–1076 (2013).
117. Adesnik, H., Bruns, W., Taniguchi, H., Huang, Z. J. & Scanziani, M. A neural circuit for spatial summation in visual cortex. *Nature* **490**, 226–231 (2012).
118. Gentet, L. J. et al. Unique functional properties of somatostatin-expressing GABAergic neurons in mouse barrel cortex. *Nat. Neurosci.* **15**, 607–612 (2012).
119. Lee, S., Kruglikov, I., Huang, Z. J., Fishell, G. & Rudy, B. A disinhibitory circuit mediates motor integration in the somatosensory cortex. *Nat. Neurosci.* **16**, 1662–1670 (2013).
120. Olsen, S. R., Bortone, D. S., Adesnik, H. & Scanziani, M. Gain control by layer six in cortical circuits of vision. *Nature* **483**, 47–52 (2012).
121. Wilson, N. R., Runyan, C. A., Wang, F. L. & Sur, M. Division and subtraction by distinct cortical inhibitory networks in vivo. *Nat. Biotechnol.* **488**, 343–348 (2012).
122. Xue, M., Atallah, B. V. & Scanziani, M. Equalizing excitation-inhibition ratios across visual cortical neurons. *Nature* **511**, 596–600 (2014).
123. Ito-Ishida, A., Ure, K., Chen, H., Swann, J. W. & Zoghbi, H. Y. Loss of MeCP2 in parvalbumin- and somatostatin-expressing neurons in mice leads to distinct Rett syndrome-like phenotypes. *Neuron* **88**, 651–658 (2015).
124. Krishnan, K. et al. MeCP2 regulates the timing of critical period plasticity that shapes functional connectivity in primary visual cortex. *Proc. Natl Acad. Sci. USA* **112**, E4782–E4791 (2015).
125. Mierau, S. B., Patrizi, A., Hensch, T. K. & Fagioli, M. Cell-specific regulation of N-methyl-D-aspartate receptor maturation by MeCP2 in cortical circuits. *Biol. Psychiatry* **79**, 746–754 (2016).
126. He, L. J. et al. Conditional deletion of *Mecp2* in parvalbumin-expressing GABAergic cells results in the absence of critical period plasticity. *Nat. Commun.* **5**, 5036 (2014).
127. Tang, X. et al. KCC2 rescues functional deficits in human neurons derived from patients with Rett syndrome. *Proc. Natl Acad. Sci. USA* **113**, 751–756 (2016).
128. Blaesse, P., Airaksinen, M. S., Rivera, C. & Kaila, K. Cation-chloride cotransporters and neuronal function. *Neuron* **61**, 820–838 (2009).
129. Kaila, K., Price, T. J., Payne, J. A., Puskarjov, M. & Voipio, J. Cation-chloride cotransporters in neuronal development, plasticity and disease. *Nat. Rev. Neurosci.* **15**, 637–654 (2014).
130. Ben-Ari, Y. NKCC1 chloride importer antagonists attenuate many neurological and psychiatric disorders. *Trends Neurosci.* **40**, 536–554 (2017).
131. He, Q. E., Nomura, T., Xu, J. & Contractor, A. The developmental switch in GABA polarity is delayed in fragile X mice. *Eur. J. Neurosci.* **34**, 446–450 (2014).
132. Ben-Ari, Y. et al. Refuting the challenges of the developmental shift of polarity of GABA actions: GABA more exciting than ever! *Front. Cell. Neurosci.* **6**, 35 (2012).
133. Duarte, S. T. et al. Abnormal expression of cerebrospinal fluid cation chloride cotransporters in patients with Rett syndrome. *PLoS ONE* **8**, e68851 (2013).
134. Castro, J. et al. Functional recovery with recombinant human IGF1 treatment in a mouse model of Rett Syndrome. *Proc. Natl Acad. Sci. USA* **111**, 9941–9946 (2014).
135. Kelsch, W. et al. Insulin-like growth factor 1 and a cytosolic tyrosine kinase activate chloride outward transport during maturation of hippocampal neurons. *Eur. J. Neurosci.* **21**, 8359–8347 (2001).
136. Ko, H. et al. The emergence of functional microcircuits in visual cortex. *Nature* **496**, 96–100 (2013).
137. Espinosa, J. S. & Stryker, M. P. Development and plasticity of the primary visual cortex. *Neuron* **75**, 230–249 (2012).
138. Chen, S. X., Kim, A. N., Peters, A. J. & Komiyama, T. Subtype-specific plasticity of inhibitory circuits in motor cortex during motor learning. *Nat. Neurosci.* **18**, 1109–1115 (2015).
139. Cichon, J. & Gan, W.-B. Branch-specific dendritic Ca²⁺ spikes cause persistent synaptic plasticity. *Nature* **520**, 180–185 (2015).
140. Hayashi-Takagi, A. et al. Labelling and optical erasure of synaptic memory traces in the motor cortex. *Nature* **525**, 333–338 (2015).
141. Turrigiano, G. G. & Nelson, S. B. Homeostatic plasticity in the developing nervous system. *Nat. Rev. Neurosci.* **5**, 97–107 (2004).
142. Sur, M., Nagakura, I., Chen, N. & Sugihara, H. Mechanisms of plasticity in the developing and adult visual cortex. *Prog. Brain Res.* **207**, 243–254 (2013).
143. Hong, E. J., McCord, A. E. & Greenberg, M. E. A. Biological function for the neuronal activity-dependent component of *Bdnf* transcription in the development of cortical inhibition. *Neuron* **60**, 610–624 (2008).

144. Yee, A. X., Hsu, Y.-T. & Chen, L. A metaplasticity view of the interaction between homeostatic and Hebbian plasticity. *Philos. Trans. R. Soc. Lond. B Biol. Sci.* **372**, 20160155 (2017).
145. Mullins, C., Fishell, G. & Tsien, R. W. Unifying views of autism spectrum disorders: a consideration of autoregulatory feedback loops. *Neuron* **89**, 1131–1156 (2016).
146. Ebert, D. H. & Greenberg, M. E. Activity-dependent neuronal signalling and autism spectrum disorder. *Nature* **493**, 327–337 (2013).
147. Blackman, M. P., Djukic, B., Nelson, S. B. & Turrigiano, G. G. A critical and cell-autonomous role for MeCP2 in synaptic scaling up. *J. Neurosci.* **32**, 13529–13536 (2012).
148. Qiu, Z. et al. The Rett syndrome protein MeCP2 regulates synaptic scaling. *J. Neurosci.* **32**, 989–994 (2012).
149. Zhong, X., Li, H. & Chang, Q. MeCP2 phosphorylation is required for modulating synaptic scaling through mGluR5. *J. Neurosci.* **32**, 12841–12847 (2012).
150. Noutel, J., Hong, Y. K., Leu, B., Kang, E. & Chen, C. Experience-dependent retinogeniculate synapse remodeling is abnormal in MeCP2-deficient mice. *Neuron* **70**, 35–42 (2011).
151. Tropea, D., Van Wart, A. & Sur, M. Molecular mechanisms of experience-dependent plasticity in visual cortex. *Philos. Trans. R. Soc. Lond. B Biol. Sci.* **364**, 341–355 (2009).
152. Brown, K. et al. The molecular basis of variable phenotypic severity among common missense mutations causing Rett syndrome. *Hum. Mol. Genet.* **25**, 558–570 (2016).
153. Samaco, R. C. et al. *Crh* and *Oprm1* mediate anxiety-related behavior and social approach in a mouse model of MECP2 duplication syndrome. *Nat. Genet.* **44**, 206–211 (2012).
154. Sztainberg, Y. et al. Reversal of phenotypes in MECP2 duplication mice using genetic rescue or antisense oligonucleotides. *Nature* **528**, 123–126 (2015).
155. Van Esch, H. et al. Duplication of the MECP2 region is a frequent cause of severe mental retardation and progressive neurological symptoms in males. *Am. J. Hum. Genet.* **77**, 442–453 (2005).
156. Samaco, R. C. et al. Female *Mecp2*^{+/−} mice display robust behavioral deficits on two different genetic backgrounds providing a framework for pre-clinical studies. *Hum. Mol. Genet.* **22**, 96–109 (2013).
157. Goffin, D. et al. Rett syndrome mutation MeCP2 T158A disrupts DNA binding, protein stability and ERP responses. *Nat. Neurosci.* **15**, 274–283 (2011).
158. Lamonica, J. M. et al. Elevating expression of MeCP2 T158M rescues DNA binding and Rett syndrome-like phenotypes. *J. Clin. Invest.* **127**, 1889–1904 (2017).
159. Veeraragavan, S. et al. Loss of MeCP2 in the rat models regression, impaired sociability and transcriptional deficits of Rett syndrome. *Hum. Mol. Genet.* **25**, 3284–3302 (2016).
160. Chen, Y. et al. Modeling Rett syndrome using TALEN-edited MECP2 mutant cynomolgus monkeys. *Cell* **169**, 945–955.e10 (2017).
161. Khwaja, O. S. et al. Safety, pharmacokinetics, and preliminary assessment of efficacy of mecasermin (recombinant human IGF-1) for the treatment of Rett syndrome. *Proc. Natl Acad. Sci. USA* **111**, 4596–4601 (2014).
162. Li, W. & Pozzo-Miller, L. BDNF deregulation in Rett syndrome. *Neuropharmacology* **76**, 737–746 (2014).
163. Katz, D. M. et al. Rett syndrome: crossing the threshold to clinical translation. *Trends Neurosci.* **39**, 100–113 (2016).
164. Chang, Q., Khare, G., Dani, V., Nelson, S. & Jaenisch, R. The disease progression of *Mecp2* mutant mice is affected by the level of BDNF expression. *Neuron* **49**, 341–348 (2006).
165. US National Library of Medicine. *ClinicalTrials.gov* <https://clinicaltrials.gov/ct2/show/NCT02061137> (2014).
166. US National Library of Medicine. *ClinicalTrials.gov* <https://clinicaltrials.gov/ct2/show/NCT02153723> (2014).
167. El-Husseini, A. E. D., Schnell, E., Chetkovich, D. M., Nicoll, R. A. & Brecht, D. S. PSD-95 involvement in maturation of excitatory synapses. *Acta Crystallogr. B* **290**, 1364–1368 (2000).
168. Sheng, M. & Kim, M. J. Postsynaptic signaling and plasticity mechanisms. *Acta Crystallogr. B* **298**, 776–780 (2002).
169. Yoshii, A. & Constantine-Paton, M. BDNF induces transport of PSD-95 to dendrites through PI3K-AKT signaling after NMDA receptor activation. *Nat. Neurosci.* **10**, 702–711 (2007).
170. Zheng, W. H. & Quirion, R. Comparative signaling pathways of insulin-like growth factor-1 and brain-derived neurotrophic factor in hippocampal neurons and the role of the PI3 kinase pathway in cell survival. *J. Neurochem.* **89**, 844–852 (2004).
171. Chao, H.-T., Zoghbi, H. Y. & Rosenmund, C. MeCP2 controls excitatory synaptic strength by regulating glutamatergic synapse number. *Neuron* **56**, 58–65 (2007).
172. Farra, N. et al. Rett syndrome induced pluripotent stem cell-derived neurons reveal novel neurophysiological alterations. *Mol. Psychiatry* **17**, 1261–1271 (2012).
173. Marchetto, M. C. N. et al. A model for neural development and treatment of Rett syndrome using human induced pluripotent stem cells. *Cell* **143**, 527–539 (2010).
174. Williams, E. C. et al. Mutant astrocytes differentiated from Rett syndrome patients-specific iPSCs have adverse effects on wild-type neurons. *Hum. Mol. Genet.* **23**, 2968–2980 (2014).
175. Yazdani, M. et al. Disease modeling using embryonic stem cells: MeCP2 regulates nuclear size and RNA synthesis in neurons. *Stem Cells* **30**, 2128–2139 (2012).
176. Lancaster, M. A. et al. Cerebral organoids model human brain development and microcephaly. *Nature* **501**, 373–379 (2012).
177. Quadrato, G. et al. Cell diversity and network dynamics in photosensitive human brain organoids. *Nature* **545**, 48–53 (2017).
178. Di Lullo, E. & Kriegstein, A. R. The use of brain organoids to investigate neural development and disease. *Nat. Rev. Neurosci.* **18**, 573–584 (2017).
179. Sahin, M. & Sur, M. Genes, circuits, and precision therapies for autism and related neurodevelopmental disorders. *Science* **350**, aab3897 (2015).
180. Luikenhuis, S., Giacometti, E., Beard, C. F. & Jaenisch, R. Expression of MeCP2 in postmitotic neurons rescues Rett syndrome in mice. *Proc. Natl Acad. Sci. USA* **101**, 6033–6038 (2004).
181. Hao, S. et al. Forniceal deep brain stimulation rescues hippocampal memory in Rett syndrome mice. *Nature* **526**, 430–434 (2015).
182. Lu, H. et al. Loss and gain of MeCP2 cause similar hippocampal circuit dysfunction that is rescued by deep brain stimulation in a Rett syndrome mouse model. *Neuron* **91**, 739–747 (2016).
183. Chiken, S. & Nambu, A. Mechanism of deep brain stimulation: inhibition, excitation, or disruption? *Neuroscientist* **22**, 313–322 (2016).
184. Garg, S. K. et al. Systemic delivery of MeCP2 rescues behavioral and cellular deficits in female mouse models of Rett syndrome. *J. Neurosci.* **33**, 13612–13620 (2013).
185. Brendel, C. et al. Readthrough of nonsense mutations in Rett syndrome: evaluation of novel aminoglycosides and generation of a new mouse model. *J. Mol. Med.* **89**, 389–398 (2011).
186. Vecsler, M. et al. Ex vivo treatment with a novel synthetic aminoglycoside NB54 in primary fibroblasts from Rett syndrome patients suppresses MECP2 nonsense mutations. *PLoS ONE* **6**, e20733 (2011).
187. Sripathy, S. et al. Screen for reactivation of MeCP2 on the inactive X chromosome identifies the BMP/TGF- β superfamily as a regulator of XIST expression. *Proc. Natl Acad. Sci. USA* **114**, 1619–1624 (2017).
188. Carrette, L. L. G. et al. A mixed modality approach towards Xi reactivation for Rett syndrome and other X-linked disorders. *Proc. Natl Acad. Sci. USA* **115**, E668–E675 (2018).
189. Sinnamon, J. R. et al. Site-directed RNA repair of endogenous *Mecp2* RNA in neurons. *Proc. Natl Acad. Sci. USA* **114**, 201715320 (2017).

This study demonstrates that site-directed RNA editing is able to repair, at the mRNA level, a RTT-causing mutation affecting the mouse MeCP2 MBD.

Acknowledgements

The authors thank H. Tsang for thoughtful discussions and critical reading of the manuscript. The authors apologize to colleagues whose research was not cited owing to the broad scope and space limitations of this Review. The authors' research is supported by a Human Frontier Science Program Long-Term Fellowship (J.P.K.I.) and by US National Institutes of Health grants MH085802 and EY007023, and the Simons Foundation Autism Research Initiative through the Simons Center for the Social Brain (M.S.).

Author contributions

J.P.K.I., N.M. and M.S. researched data for the article, contributed substantially to the discussion of content, wrote the article and reviewed and edited the manuscript before submission.

Competing interests

The authors declare no competing interests.

Publisher's note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Reviewer information

Nature Reviews Neuroscience thanks T. Pizzorusso and the other anonymous reviewer(s) for their contribution to the peer review of this work.